

Introduction and spread of non-native parasites with *Silurus glanis* L. (Teleostei: Siluridae) in UK fisheries

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Abstract

Despite growing concern of the ecological risks posed by the European catfish *Silurus glanis* L. in freshwater fisheries, little information exists on the parasite fauna of this silurid catfish in Britain. Parasitological examinations of released *S. glanis* from four still-water fisheries in England revealed the presence of *Thaparocleidus vistulensis* (Siwak, 1932) and *Ergasilus sieboldi* (Nordmann, 1832), both non-native parasites, the latter known to be an important fish pathogen. This represents the first record of *T. vistulensis* from British freshwater fish. The human-assisted movement of *S. glanis* between UK recreational still-water fisheries provides a clear avenue for the introduction and spread of non-native parasites.

Introduction

An inherent and persistent risk associated with fish introductions is the transmission of their parasitic fauna to native host fishes (Kennedy, 1994; Kirk, 2003). These naïve hosts may be highly susceptible to infection where there has been a lack of host–parasite co-evolution, which may result in low natural immunity, altered disease dynamics and poor anti-parasite behaviour (Taraschewski, 2006; Kelly *et al.*, 2009). Transmission of non-native parasites to new geographical localities has already resulted in serious and irreversible effects in some species, such as the detrimental effect of *Anguillicoloides crassus* (Kuwahara, Niimi *et al.*, 1974) on the European eel *Anguilla anguilla* L. (Székely *et al.*, 2009). The global spread of parasites with invading hosts is therefore a major cause of disease emergence and holds important implications for native aquatic environments (Gozlan *et al.*, 2005, 2009; Perkins *et al.*, 2008; Peeler *et al.*, 2011).

In England and Wales the European catfish *Silurus glanis* L. has been widely introduced for enhancing the performance of recreational lake fisheries (Britton *et al.*, 2010). Many of these introductions have been completed outside of relevant regulatory frameworks and so have not been subjected to risk assessment processes prior to their release (Hickley & Chare, 2004; Copp *et al.*, 2005, 2009). In order to better understand the ecological risks associated with this species, research has been conducted to assess the invasiveness of this fish in the UK. Most of this work has focused on colonization potential under current and future climatic conditions (Britton *et al.*, 2007, 2010; Copp *et al.*, 2009). However, according to Copp *et al.* (2009), virtually all aspects of the environmental biology of *S. glanis* require further study. This includes disease risk, as to date little attention has been given to the parasite fauna of this fish in Britain, despite awareness that it may serve as host for a wide range of parasite species (Copp *et al.*, 2009). In an effort to address this knowledge gap, the present study describes the results of parasitological investigations of *S. glanis* from freshwater fisheries.

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Materials and methods

Between 2009 and 2011, five specimens of *S. glanis* (50–170 cm) were examined from four different still-water fisheries in England. These fish were made available as part of Environment Agency investigations into the colonization potential and invasion biology of *S. glanis* in inland waters in England and Wales. These sites were located in Staffordshire, Hampshire, Kent and Essex (precise locations withheld for confidentiality).

All fish were captured by means of seine netting and transported alive to holding facilities at the Environment Agency, Brampton. Fish were killed by lethal anaesthesia (benzocaine solution 5% w/v) and examined for external and internal parasites, using low- and high-power light microscopy. The identity, site of attachment and approximate intensity of any parasites that were encountered were recorded.

Results and discussion

The ancyrocephalid monogenean parasite, *Thaparocleidus vistulensis* (Siwak, 1932) was recorded on the gills of all five fish (intensity range 1–35 per gill arch). Identification was confirmed from examination of the copulatory complex (fig. 1a) and the haptor sclerites (fig. 1b and c).

Other parasite findings included light infections of *Camallanus lacustris* (Zoega, 1776) in the intestinal tract (mean intensity, 1), *Argulus foliaceus* (L.) on the skin (mean

intensity, 1), *Diplostomum* spp. in the eye (intensity range 1–5), *Trypanosoma* spp. in the kidney and *Ergasilus sieboldi* (Nordmann, 1832) on the gill lamellae (intensity range 1–10 per gill arch).

These studies provide the first record of *T. vistulensis* in the UK. This parasite is a specialist of siluriform fishes. Voucher specimens of *T. vistulensis* have been deposited in the parasitic worm collection of the Natural History Museum, London (NHMUK 2011.10.27.1-3). *Ergasilus sieboldi* is also a non-native parasite, but may infect a wide range of fish species (Fryer, 1969).

Thaparocleidus spp. have been recorded from much of Asia and Europe (Lim *et al.*, 2001; Moravec, 2001; Galli *et al.*, 2003; Copp *et al.*, 2009). Specific records for *T. vistulensis* include Italy, the Czech Republic, the Slovak Republic and Poland (Siwak, 1932; Moravec, 2001; Paladini *et al.*, 2008), although these are likely to reflect detection effort rather than the true extent of distribution. Limited information exists on the pathogenicity of *T. vistulensis* and the pathology of this parasite has not been described. Blanc (1997) listed a number of ancyrocephalid Monogenea in a table of introduced fish pathogens. However, the pathogenic importance of these species was not detailed.

Despite the examination of only a small number of fish, the recording of *T. vistulensis* confirms the potential for introduced fish to concomitantly introduce their parasitic fauna into areas outside of their natural range. This finding closely follows the detection of the ancyrocephalid

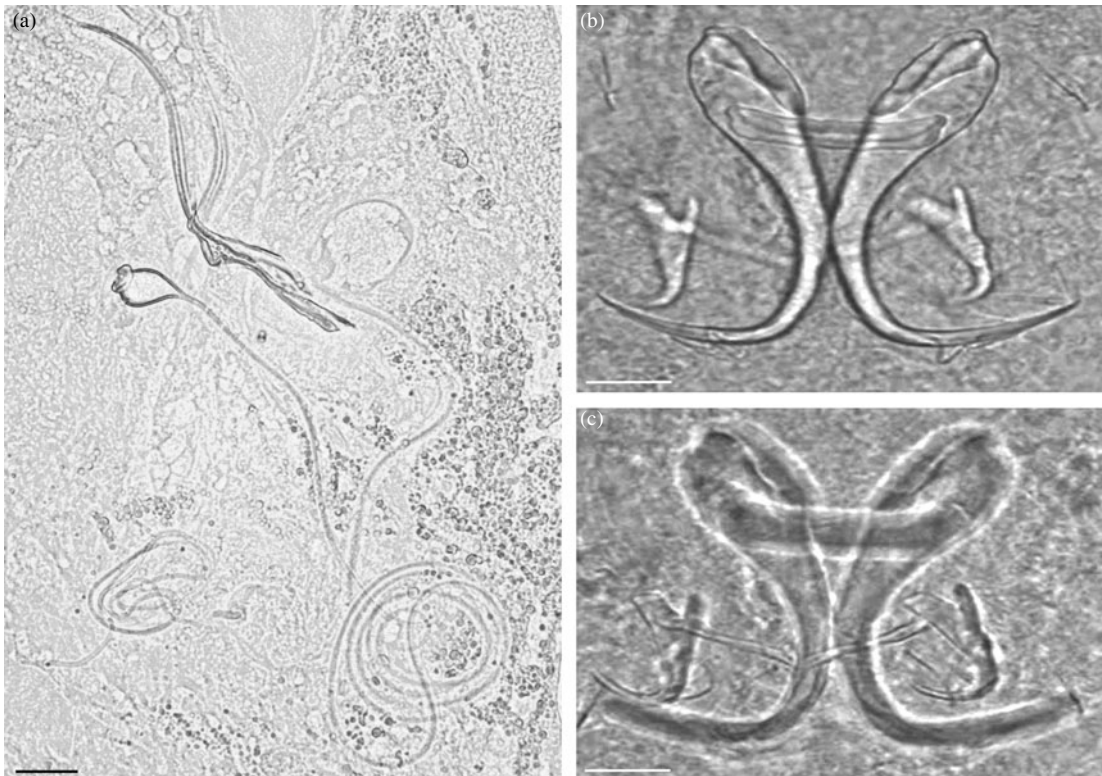


Fig. 1. Copulatory complex (a) and haptor sclerites (b, c) of *Thaparocleidus vistulensis* (Siwak, 1932). Scale bar = 20 μ m.

monogenean *Onchocleidus dispar* (Müller, 1936) in pumpkinseed *Lepomis gibbosus* (L.) introduced into the UK (Hockley *et al.*, 2011). Andrews & Chubb (1984) recorded *Proteocephalus osculatus* (Goeze, 1782), a common parasite of catfish in Russia, from *S. glanis* imported to a fish farm in Yorkshire, England. Although the fate of these fish was not detailed, it can be assumed that parasite colonization was prevented as these fish underwent antihelminthic treatment.

Many non-native monogeneans have been recorded in Europe following fish translocations (Johnson & Jensen, 1991; Moravec, 2001; Galli *et al.*, 2003). Although the potential for disease surrounds any parasite introduction (Kennedy, 1994), the risk posed by these monogeneans may be limited due to their purported strict host specificity. *Thaparocleidus vistulensis* is restricted to freshwater siluriforms that are naturally absent from the UK fish fauna (Lim *et al.*, 2001; Davies *et al.*, 2004; Paladini *et al.*, 2008).

However, the simultaneous detection of *E. sieboldi* in the examined fish highlights a potential disease risk to native host species. The high reproductive rate, direct life-cycle and low host specificity of many ergasilid parasites have led to their rapid colonization and spread. *Ergasilus sieboldi* has a predilection for large fish and has been the cause of mortality in a range of fish species in still-water fisheries (Alston & Lewis, 1994; Tildesley, 2008). The spread of this parasite with wels catfish represents an additional disease risk, as fishery managers and anglers have a propensity to stock large specimens of *S. glanis* for enhancing lake fisheries (Hickley & Chare, 2004; Britton *et al.*, 2007). These human-driven fish movements are the result of the commoditization of such valuable fish within the UK recreational fishery sector (Hickley & Chare, 2004) and provide a dispersal pathway for the introduction and spread of fish pathogens.

The translocation of parasites with the trade in fish represents a considerable threat to aquatic biodiversity and fishery development (Copp *et al.*, 2005; Gozlan *et al.*, 2006). Examples of disease outbreaks following the introduction of non-native parasites, including a number of monogeneans, are well documented (Johnson & Jensen, 1991; Bauer *et al.*, 2002; Matsche *et al.*, 2010). Many biotic and abiotic factors influence the colonization, establishment and pathogenicity of introduced parasites (Kennedy, 1994). However, the illegal release of alien fish species into the wild (BBC, 2008), and increasing demand for large and unusual species like catfish and sturgeon, *Acipenser* spp., as sport fish (Hickley & Chare, 2004), provide clear avenues for disease transfer. The introduction of *Bothriocephalus acheilognathi* Yamaguti, 1934 with imported fathead minnow *Pimephales promelas* Rafinesque, 1820 (C. Williams, pers. obs.) and the accidental discovery of the Rosette agent *Sphaerothecum destruens* Arkush, Mendoza, Adkison et Hedrick, 2003, in the highly invasive topmouth gudgeon *Pseudorasbora parva* (Temminck et Schlegel, 1846) (Gozlan *et al.*, 2009), further illustrate these dangers. Current risk assessment frameworks may not be sufficient to prevent new parasite introductions from occurring, emphasizing the need for continued disease monitoring of high-risk fish movements and strengthening regulatory efforts to protect freshwater fisheries.

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