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**A New Species of Sucking Louse (Phthiraptera: Anoplura: Polyplacidae) from the Gray
Mouse Lemur, *Microcebus murinus*, in Madagascar**

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Hasiniaina^{5,6} and Sarah Zohdy⁷

23 **Abstract**

24 *Lemurpediculus madagascariensis* sp. nov. (Phthiraptera: Anoplura: Polyplacidae) is described
25 from the Gray Mouse lemur, *Microcebus murinus* (J. F. Miller), from Ankarafantsika National
26 Park, Madagascar. Lemurs were trapped using Sherman Live Traps and visually inspected for
27 lice, which were preserved in 90% ethanol. Adults of both sexes and the third instar nymph of
28 the new species are illustrated and distinguished from the four previously known species of
29 *Lemurpediculus*: *L. verruculosus* (Ward), *L. petterorum* Paulian, *L. claytoni* Durden, Blanco and
30 Seabolt, and *L. robbinsi* Durden, Blanco and Seabolt. It is not known if the new species of louse
31 is a vector of any pathogens or parasites.

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33 **Key Words:** Phthiraptera, Anoplura, new species, mouse lemur, Madagascar

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45 The mouse and dwarf lemurs of Madagascar (family Cheirogaleidae) are among the smallest
46 primates in the world (Zimmermann and Radespiel 2014, Lehman et al. 2016, Zohdy and Durden
47 2016). Ectoparasites of cheirogaleid lemurs are inadequately known (Blanco et al. 2013, Zohdy
48 and Durden 2016, Durden et al. 2017) and sucking lice (Phthiraptera: Anoplura) have been
49 described from only three of the more than 30 species of cheirogaleids currently recognized
50 (Hotaling et al. 2016). The three previously described species are *Lemurpediculus verruculosus*
51 (Ward), an ectoparasite of the eastern mouse lemur, *Microcebus rufus* É. Geoffroy,
52 *Lemurpediculus claytoni* Durden, Blanco and Seabolt, an ectoparasite of Sibree's dwarf lemur,
53 *Cheirogaleus sibreei* Forsyth Major, and *Lemurpediculus robbinsi*, which parasitizes Crossley's
54 dwarf lemur, *Cheirogaleus crossleyi* A. Grandidier (Durden et al. 2017). Another congeneric
55 species, *Lemurpediculus petterorum* Paulian, parasitizes a different species of lemur which was
56 stated to probably be *Lepilemur mustelinus* I. Geoffroy by Paulian (1958). Sucking lice are often
57 host-specific (Durden and Musser 1994), and because few lemur species have been sampled for
58 ectoparasites, there are undoubtedly additional undescribed species of *Lemurpediculus* associated
59 with other species of cheirogaleids. Durden et al. (2017) amended the description of the genus
60 *Lemurpediculus* to accommodate new developments in the systematics of Anoplura since the
61 genus was erected by Paulian (1958). In this paper, we describe a new species of *Lemurpediculus*
62 from the gray mouse lemur from Ankarafantsika National Park in northwestern Madagascar.

63

64 **Materials and Methods**

65 As part of a study on lemur health and communication, mouse lemurs were trapped in
66 Ankarafantsika National Park, Madagascar in Jardin Botanique A using Sherman Live Traps (H.

67 B. Sherman Traps, Inc., Tallahassee, FL) baited with banana between May and November in
68 2010 and 2011 (dry season). The study was conducted in the dry season because that is when the
69 lemurs most readily enter traps for the banana. Lemurs were removed from the traps, manually
70 restrained, and inspected for parasites by parting the fur down to the skin. Ectoparasites were
71 collected primarily from the face, ears, legs, back and tail and stored in 90% ethanol in
72 individually labeled vials. All lemurs were released at their capture site following collection of
73 data and ectoparasites. This study was approved by Madagascar National Parks
74 (N101/11/MEF/SG/DGF/DCB.SAP/SCB, N102/11/MEF/SG/DGF/DCB.SA/SCB) and the
75 Arizona State University Institutional Animal Care and Use Committee (Protocol: 10-1077R).

76 Lice were cleared in 10% potassium hydroxide for ~24 h, rinsed in distilled water,
77 transferred to 70% ethanol and then slide mounted in PVA medium (Bioquip Products, Rancho
78 Dominguez, CA). Slide-mounted lice, including specimens of previously described congeneric
79 species (in the Anoplura collections of LAD), were examined at high magnification under phase-
80 contrast using an Olympus BH-2 microscope (Olympus Corporation of the Americas, Center
81 Valley, PA) connected to an Ikegami MTV-3 video camera attachment and monitor (Ikegami
82 Electronics, Neuss, Germany). Drawings of diagnostic morphological features were made from
83 specimens examined at 100x – 400x. Specimen measurements were made using a calibrated
84 graticule fitted into a microscope eyepiece.

85 Descriptive format for the new species follows Durden et al. (2010) and names and
86 abbreviations of setae and morphological structures follow Kim and Ludwig (1978). Names of
87 setae and certain structures are spelled out in full at first mention (with the abbreviation listed
88 parenthetically) and then abbreviated when subsequently mentioned. The holotype male, allotype
89 female and paratype third instar nymph of the new species are deposited in the U.S. National

90 Museum of Natural History (NMNH) (Smithsonian Institution), Department of Entomology,
91 Washington DC.

92 Lemur taxonomy and common names used in this paper follow Groves (2005) and
93 Hotaling et al. (2016).

94

95 **Results**

96 We trapped a total of 107 *M. murinus*. The entire ectoparasite faunas of these mouse lemurs will
97 be reported in a separate paper.

98

99 *Lemurpediculus madagascariensis* **sp. nov.** (Figs. 1-3)

100 **Male (Fig. 1A,B,C)**

101 Total body length: 1.07–1.13 mm; mean, 1.10 mm (n=5). Head, thorax and abdomen lightly
102 sclerotized.

103 Head: More heavily sclerotized along anterior dorsal margin and antero-laterally adjacent
104 to first antennal segment; longer than broad with squarish anterior margin. One long Dorsal
105 Principal Head Seta (DPHS), one small Dorsal Accessory Head Seta (DAcHS) anteromedial to
106 DPHS, one Dorsal Posterior Central Head Seta (DPoCHS), 2-3 Dorsal Preantennal Head Setae
107 (DPaHS), two Sutural Head Setae (SHS), three Dorsal Marginal Head Setae (DMHS), 3-4 Apical
108 Head Setae (ApHS) and one Ventral Preantennal Head Seta (VPaHS) on each side. Antennae
109 five-segmented with basal segment slightly wider than long and distinctly broader than second
110 segment; fourth segment slightly extended posterolaterally.

111 Thorax: Much longer than wide, slightly wider than head. Thoracic sternal plate (Fig. 1B)
112 lightly sclerotized, with narrow anterior extension and broadly curved lateral margins; tiny

113 sclerite bearing two long setae immediately posterior to thoracic sternal plate. Dorsal Principal
114 Thoracic Seta (DPTS) mean length 0.13 mm (range, 0.12-0.14 mm, n=8), with adjacent small
115 Dorsal Mesothoracic Seta (DMsS) on each side; mesothoracic spiracle mean maximum diameter
116 0.025 mm (range, 0.023-0.027 mm, n=8). Legs with subtriangular coxae; forelegs each
117 terminating in small tibio-tarsal claw; mid and hindlegs subequal in size, each terminating in
118 large, robust tibio-tarsal claw.

119 Abdomen: Wider than thorax with six annulated spiracles on each side. Paratergal plates,
120 tergites and sternites absent. One row of two long Dorsal Central Abdominal Setae (DCAS)
121 anteriorly, followed by five rows of 4-6 long DCAS and then two rows of two shorter DCAS.
122 Six Dorsal Lateral Abdominal Setae (DLAS) on each side, each adjacent to corresponding
123 spiracle; DLAS 1-5 each with adjacent small seta; DLAS 1 and adjacent small seta both inserted
124 on small ridge. DLAS 6 borne on small sclerite and distinctly longer than other DLAS and
125 extending away from abdomen; five rows of four long Ventral Central Abdominal Setae
126 (VCAS); VCAS in most posterior row slightly shorter than other VCAS. One posterior Ventral
127 Lateral Abdominal Seta (VLAS) on each side adjacent to corresponding DLAS and most
128 posterior spiracle. ~10 tiny to small dorsal setae near posterior apex of abdomen.

129 Genitalia (Figs 1A,C): Subgenital plate (Fig. 1A) well sclerotized, somewhat urn-shaped
130 with bulging medio-lateral margins and small antero-lateral extensions. Basal apodeme longer
131 than parameres and other genitalic components combined, slightly expanded posteriorly into two
132 paddle-shaped plates on each side; C-shaped anterior endomere with posteriorly converging
133 arms; anteriorly acuminate aedeagal sclerite located inside anterior endomere; posteriorly
134 acuminate central endomere bordered laterally by one broad accessory sclerite on each side;

135 parameres broad anteriorly and tapering posteriorly to rounded apex; pseudopenis relatively
136 small but extending posteriorly beyond apices of parameres and terminating in acute apex.

137

138 **Female (Fig. 2A,B,C)**

139 Body length: 1.32-1.50 mm; mean, 1.43 mm (n=6). Head, thorax and abdomen as in male unless
140 indicated otherwise.

141 Head: Slightly wider than in male.

142 Thorax: Mesothoracic spiracle mean maximum diameter 0.0275 mm (range, 0.0250-
143 0.0283, n=6).

144 Abdomen: Dorsally with eight rows of four long DCAS anteriorly followed by one row
145 of six slightly shorter DCAS and one row six small Tergal Abdominal Setae (TeAS) inserted on
146 broad, curved tergite immediately posterior to subgenital plate. One row of one DLAS on each
147 side anteriorly followed by six rows of two DLAS on each side and then one very long DLAS
148 borne on small sclerite posterior to last spiracle. One row of two long VCAS anteriorly followed
149 by five rows of four VCAS. One very long VLAS posteriorly, associated with last DLAS and
150 most posterior spiracle.

151 Genitalia (Fig. 2C): Subgenital plate broadly rounded anteriorly and extending
152 posteriorly to broad apex, with small, distinct lateral lacuna on each side; each lacuna with four
153 small setae inserted anteriorly; three small setae inserted on each side of subgenital plate near
154 postero-lateral margins. Vulvar fimbriae distinct and extensive collectively forming a V shape;
155 gonopods VIII and IX indistinct and with ~13 setae attached to each gonopod VIII and two
156 slightly larger setae attached to each gonopod IX; gonopod setae collectively forming postero-

157 lateral fan-like patches. Curved subterminal transverse sclerite with small anterior apex situated
158 between gonopods IX. Three small terminal setae ventrally on each side of genital opening.

159

160 **Third Instar Nymph (Fig. 3)**

161 Body length: 1.00-1.25 mm; mean, 1.11 mm (n=5).

162 Head: Shape as in male but with slightly more rounded anterior margin. One fairly long
163 DPTS and one adjacent small DAcHS, one DPoCHS, two DMHS, two SHS, 3-4 ApHS and one
164 VP aHS on each side. Antennae approximately as in male.

165 Thorax: Slightly wider than head, much longer than wide; one long DPTS (mean length,
166 0.125 mm, range, 0.110-0.129 mm, n=4) adjacent to mesothoracic spiracle (mean maximum
167 diameter, 0.025 mm, range, 0.020-0.028, n=4) on each side. Foreleg coxae subtriangular; mid
168 and hind coxae more irregular; forelegs each terminating in small tibio-tarsal claw; mid and
169 hindlegs subequal in size, each terminating in large, robust tibio-tarsal claw.

170 Abdomen: Wider than thorax with eight rows of two DCAS and nine rows of VCAS.
171 Eight DLAS on each side; DLAS 2-7 each with accompanying spiracle; DLAS 2 with adjacent
172 small accessory seta, both borne on small protuberance; two additional small setae adjacent to
173 DLAS 2 on each side; one additional small accessory seta on each side adjacent to each of DLAS
174 3-6; DLAS 7 and 8 both long, extending from postero-lateral abdomen and each associated with
175 one VLAS on each side.

176 **HOLOTYPE** ♂ ex *Microcebus murinus* (J. F. Miller) (gray mouse lemur) (male, Animal
177 25-09), MADAGASCAR: Boeny Region, Ankarafantsika National Park, Jardin Botanique A
178 (46°48' E, 16°19' S), elevation 190 m, 17 October 2010, Coll: Sharon Kessler and Alida I. F.
179 Hasiniaina. Deposited in NMNH (accession barcode, USNMENT00981907).

180 **ALLOTYPE** ♀ ex *M. murinus*, same data as Holotype except (male, Animal 82-10) and
181 13 Nov. 2011. Deposited in NMNH (accession barcode, USNMENT00981908).

182 **PARATYPES** One nymph (third instar) same data as Holotype except (male, Animal 25-
183 10), 15 October 2010 (accession barcode, USNMENT00981909); 2♂, 2♀, 2 nymphs (third
184 instars) same data as Holotype except different individual lemurs and various dates in 2010 and
185 2011; deposited in Georgia Southern University Insect Collection (1♂, 1♀) (accession no. L-
186 3813) and Anoplura Collection of L. A. Durden (1♂, 1♀).

187 **ETYMOLOGY:** This species is named for the faunistically unique island of Madagascar where
188 both the louse and its host co-occur.

189 For comparative purposes, the female subgenital plates and associated structures, for the
190 four previously described congeneric species, *L. petterorum*, *L. claytoni*, *L. verruculosus*, and *L.*
191 *robbinsi*, are illustrated in Fig. 4.

192

193 **Discussion**

194 **Males**

195 Males of *Lemurpediculus* spp. can easily be separated by examination of the genitalia in
196 cleared slide-mounted specimens. In *L. petterorum* males, the parameres are about equal in
197 length to the basal apodeme (shown in Paulian 1958, Fig. 1B), whereas *L. claytoni*, *L. robbinsi*,
198 and *L. verruculosus*, the parameres are much shorter than the basal apodeme. The shape of the
199 parameres and the presence or absence of genitalic endomeres and accessory sclerites can be
200 used to separate these four species. The parameres have slightly concave medio-lateral margins
201 in *L. claytoni* (shown in Durden et al. 2017, Fig. 3B) and distinctly rounded convex medio-lateral
202 margins in both *L. verruculosus* (shown in Durden et al. 2010, Fig. 3) and *L. robbinsi* (shown in

203 Durden et al. 2017, Fig. 2B). The medial margins of the parameres of *L. robbinsi* have a
204 distinctly rounded bulge (shown in Durden et al. 2017, Fig. 2B) which is absent in *L.*
205 *verruculosus* (shown in Durden et al. 2010, Fig. 3). Further, the pseudopenis extends well
206 beyond the posterior apices of the parameres in *L. robbinsi* (shown in Durden et al. 2017, Fig.
207 2B) but just barely beyond the apices in *L. verruculosus* (shown in Durden et al. 2010, Fig. 3).
208 The male genitalia of *L. madagascariensis* sp. nov. (Fig. 1C) have more acute anterior paramere
209 margins than those of the other species in the genus and have additional adjacent small plates
210 that are not present in the other species – a central endomere and a pair of lateral sclerites.
211 Externally, the thoracic sternal plate of *L. petterorum* (shown in Paulian 1958, Fig. 2B) lacks an
212 anterior projection which is clearly present in the other four species.

213 **Females**

214 Females of all five known species of *Lemurpediculus* can easily be separated based on
215 the shape of the subgenital plate which can be observed in either cleared or uncleared specimens.
216 In both *L. petterorum* and *L. madagascariensis* sp. nov., the anterior and posterior portions of the
217 subgenital plate are joined centrally and laterally and have two lateral lacunae in the anterior
218 portion (Figs. 2C, 4A), whereas the anterior and posterior sections of the subgenital plate are not
219 joined laterally in the other three species Fig. 4B-D). The two lacunae in the female subgenital
220 plate of *L. petterorum* are much larger than those in *L. madagascariensis*, collectively making up
221 almost half of the plate size in the former species (Fig. 4A), but less than 10% in the latter
222 species (Fig. 2C). Also, the thoracic sternal plate in the female of *L. petterorum* lacks the anterior
223 extension (shown in Paulian 1958, Fig. 2B) that is present in females of *L. madagascariensis* sp.
224 nov. (Fig. 2B in this paper). In females of *L. verruculosus*, the anterior portion of the subgenital
225 plate is 3-4 times larger than the posterior portion (Fig. 4C). The anterior and posterior sections

226 of the subgenital plate are subequal in size in females of *L. claytoni* (Fig. 4B) whereas the
227 anterior portion is slightly larger than the posterior portion in *L. robbinsi* (4D). Also, the shape of
228 the female subgenital plate is very different between these species (Figs. 2C, 4A-D).

229 **Nymph**

230 The third instar nymph of only one other species of *Lemurpediculus* has been described.
231 This nymphal stage of *L. verruculosus* was described and illustrated by Durden et al. (2010). The
232 third instar nymph is easily separated between these two species because *L. verruculosus* lacks
233 DLAS next to abdominal spiracles 2-5 (see Durden et al. 2010, Fig. 3) whereas *L.*
234 *madagascariensis* sp. nov. has one long DLAS next to each of these spiracles on each side (Fig.
235 2C).

236 With the description of the new species included in this paper, there are now five
237 recognized species of *Lemurpediculus*. Four of these species, *L. verruculosus*, *L. robbinsi*, *L.*
238 *claytoni*, and *L. madagasarensis* sp. nov., parasitize cheirogaleid lemurs while the host of the
239 fifth species, *L. petterorum* Paulian, was stated by Paulian (1958) to probably be *Lepilemur*
240 *mustelinus* (weasel sportive lemur) another nocturnal species which belongs to a different lemur
241 family, the Lepilemuridae. All five of these species of lice appear to be host-specific but the
242 host/s of *L. petterorum* requires verification. It would be premature to provide a dichotomous
243 identification key for known *Lemurpediculus* species because we anticipate the collection and
244 description of additional species in this genus in the future, especially considering the highly
245 diverse radiation of mouse lemur species around Madagascar (Yoder et al. 2010, 2016).

246 With few exceptions, very little is known about the potential for sucking lice of wild
247 mammals to transmit pathogens to their hosts (Durden 2001) and nothing is currently known
248 about any potential vectorial role of lice that parasitize lemurs. However, some pathogens and

249 parasites of lemurs, including certain viruses, bacteria and protozoans could feasibly be
250 transmitted by sucking lice. Future research should address the potential for blood-feeding
251 ectoparasites, including sucking lice, to transmit pathogens to lemurs, particularly in light of the
252 threatened or endangered status of many species of these primates.

253 Many authors have advocated conserving (or co-conserving) parasites of rare hosts (e.g.,
254 Durden and Keirans 1996, Whiteman and Parker 2005, Dunn et al. 2009) and we likewise
255 advocate co-conservation of mouse lemurs and their unique host-specific parasites including
256 their sucking lice.

257

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272 lice.

273

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325

326 **Footnotes**

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337

338

339 **Figure Legends**

340 **Fig. 1.** *Lemurpediculus madagascariensis* sp. nov., adult male. A: Dorsoventral view. B:
341 Thoracic sternal plate. C. Genitalia. All scale bars, 0.1 mm.

342 **Fig. 2.** *Lemurpediculus madagascariensis* sp. nov., adult female. A: Dorsoventral view. B:
343 Thoracic sternal plate. C. Genitalia. All scale bars, 0.1 mm.

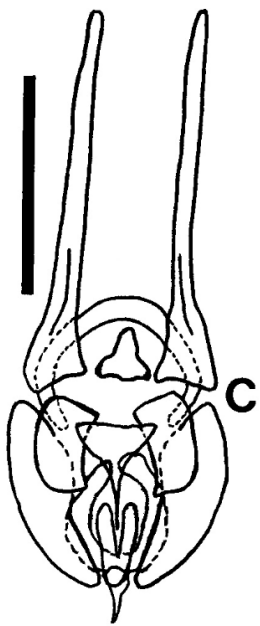
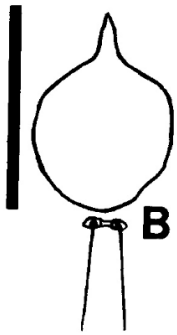
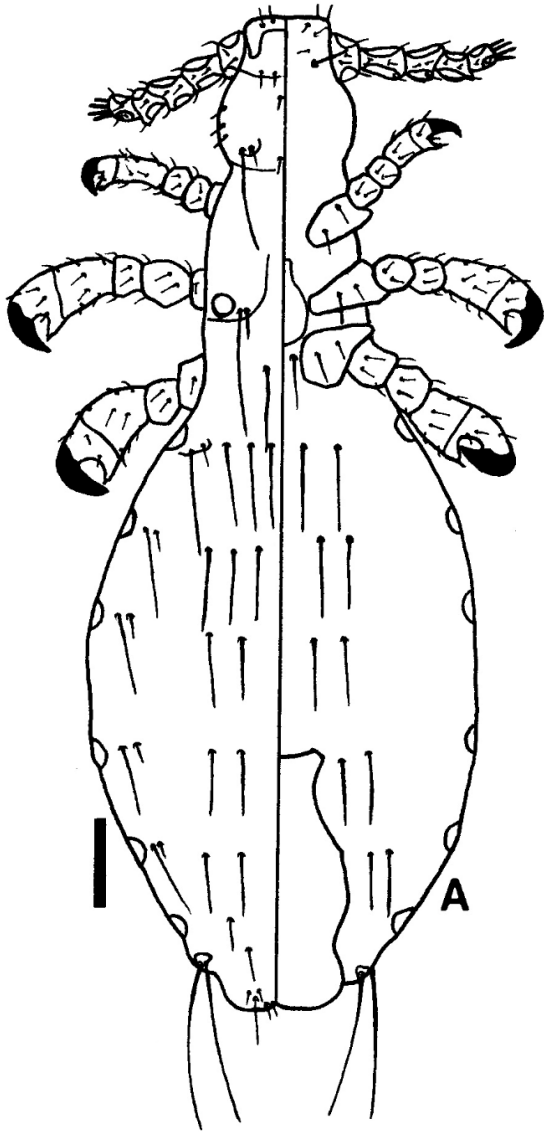
344 **Fig. 3.** *Lemurpediculus madagascariensis* sp. nov., third instar nymph: Dorsoventral view. Scale
345 bar, 0.1 mm.

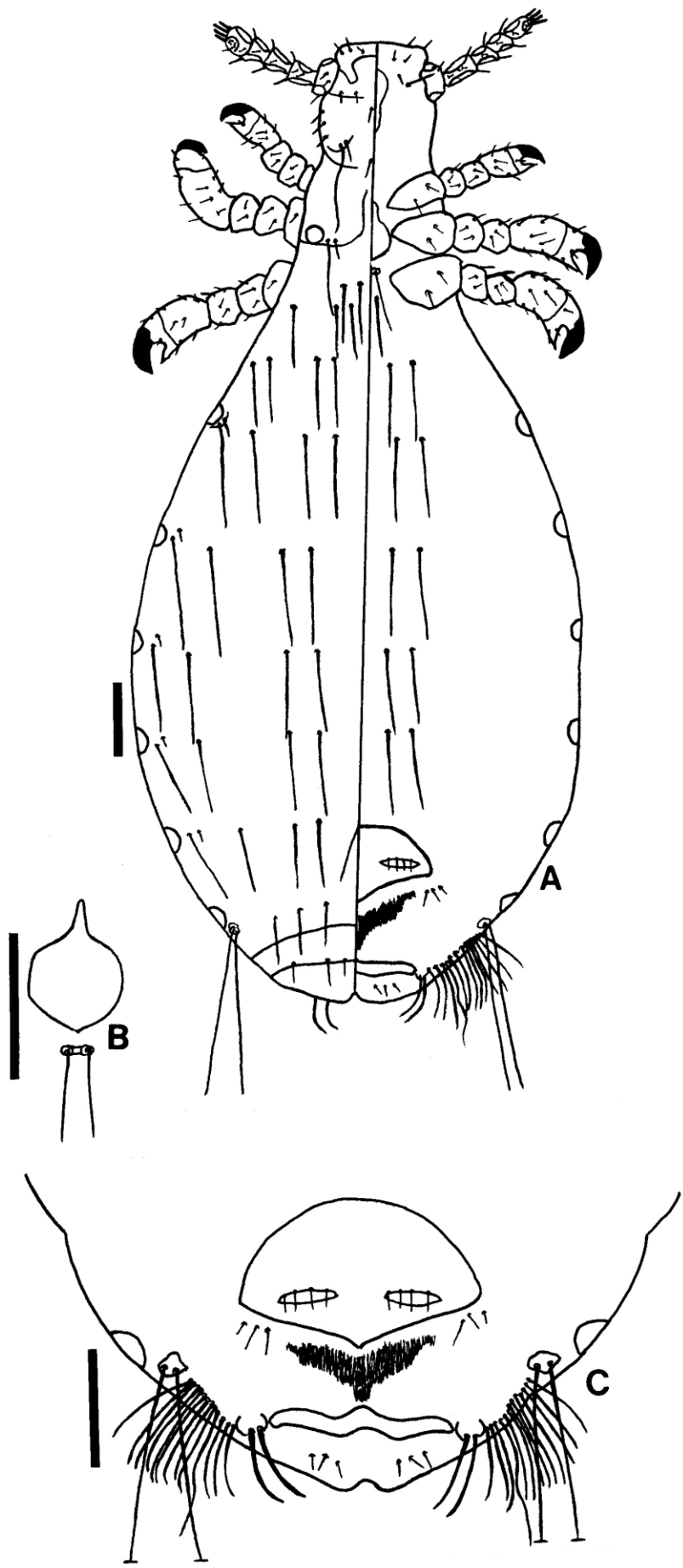
346 **Fig. 4.** Female subgenital plates, associated setae, and vulvar fimbriae of the four previously
347 described species of *Lemurpediculus*. A: *L. petterorum* Paulian ex (probably) *Lepilemur*
348 *mustelinus*, Ambatolampy District, Madagascar. B: *L. claytoni* Durden, Blanco and Seabolt ex
349 *Cheirogaleus sibreei*, Tsinjoarivo, Amabatolampy District, Madagascar. C: *L. verruculosus*
350 (Ward) ex *Microcebus rufus*, Ranomafana National Park, Madagascar. D: *L. robbinsi* Durden,
351 Blanco and Seabolt ex *Cheirogaleus crossleyi*, Tsinjoarivo, Amabatolampy District,
352 Madagascar. Scale bar = 0.05 mm.

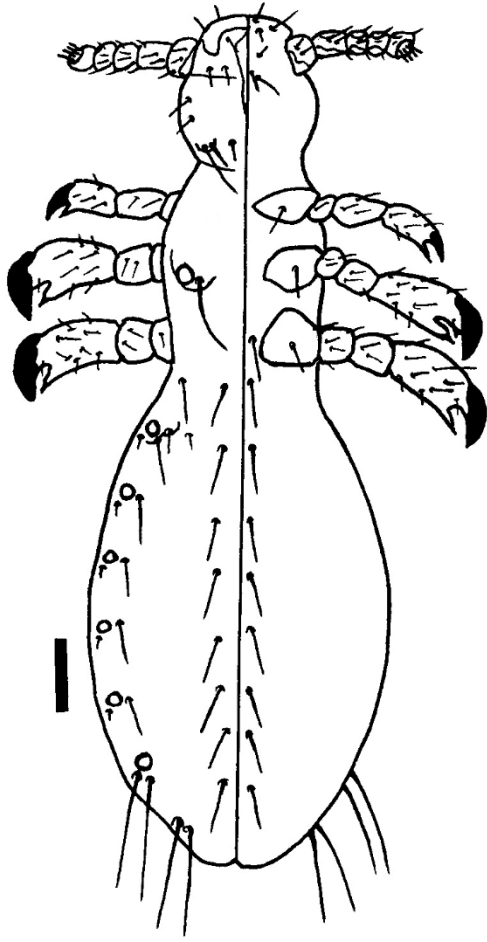
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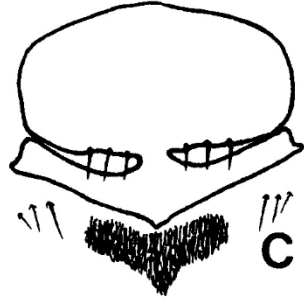
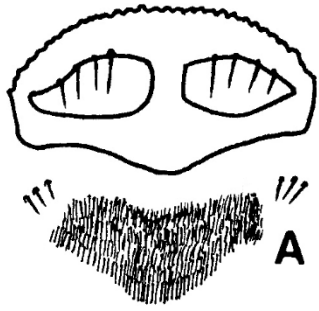
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