This is the peer reviewed version of the following article: Lowry, D., Sobel, J., Angert, A., Ashman, T.-L., Baker, R., Blackman, B., Brandvain, Y., Byers, K., Cooley, A., Coughlan, J., Dudash, M., Fenster, C., Ferris, K., Fishman, L., Friedman, J., Grossenbacher, D., Holeski, L., Ivey, C., Kay, K., Koelling, V., Kooyers, N., Murren, C., Muir, C., Nelson, T., Peterson, M., Puzey, J., Rotter, M., Seemann, J., Sexton, J., Sheth, S., Streisfeld, M., Sweigart, A., Twyford, A., Vallejo-Marín, M., Willis, J., Wright, K., Wu, C. and Yuan, Y.-W. (2019), The case for the continued use of the genus name Mimulus for all monkeyflowers. *TAXON*, 68: 617-623, which has been published in final form at https://doi.org/10.1002/tax.12122. This article may be used for non-commercial purposes in accordance with Wiley Terms and Conditions for self-archiving.

The case for the continued use of the genus name Mimulus for all monkeyflowers

Running Title: Mimulus for all monkeyflowers

- 6 David B. Lowry^{1*}, James M. Sobel², Amy L. Angert³, Tia-Lynn Ashman⁴, Robert L. Baker⁵,
- 7 Benjamin K. Blackman⁶, Yaniv Brandvain⁷, Kelsey J.R.P. Byers⁸, Arielle M. Cooley⁹, Jennifer
- 8 M. Coughlan¹⁰, Michele R. Dudash¹¹, Charles B. Fenster¹², Kathleen G. Ferris¹³, Lila Fishman¹⁴,
- 9 Jannice Friedman¹⁵, Dena L. Grossenbacher¹⁶, Liza M. Holeski¹⁷, Christopher T. Ivey¹⁸,
- 10 Kathleen M. Kay¹⁹, Vanessa A. Koelling²⁰, Nicholas J. Kooyers²¹, Courtney J. Murren²²,
- 11 Christopher D. Muir²³, Thomas C Nelson¹⁴, Megan L. Peterson²⁴, Joshua R. Puzey²⁵, Michael C.
- Rotter¹⁷, Jeffrey R. Seemann²⁶, Jason P. Sexton²⁷, Seema N. Sheth²⁸, Matthew A. Streisfeld²⁹,
- Andrea L. Sweigart³⁰, Alex D. Twyford^{31,32}, Mario Vallejo-Marín³³, John H. Willis³⁴, Kevin M.
- 14 Wright³⁵, Carrie A. Wu³⁶, Yao-Wu Yuan²⁶
- ¹Department of Plant Biology, Michigan State University, East Lansing, MI, 48824, USA
- ²Department of Biological Sciences, Binghamton University (SUNY), Binghamton, NY 13902,
- 19 USA

15 16

1 2 3

- ³Departments of Botany and Zoology, University of British Columbia, Vancouver, BC, V6T
- 21 1Z4, Canada
- ⁴Department of Biological Sciences, University of Pittsburgh, Pittsburgh, PA, 15260, USA
- ⁵Department of Biology, Miami University, Oxford, OH, 45056, USA
- ⁶Department of Plant & Microbial Biology, University of California, Berkeley, Berkeley, CA,
- 25 94720, USA
- ⁷Department of Plant & Microbial Biology, University of Minnesota, St. Paul, MN, 55108, USA
- ⁸Department of Zoology, University of Cambridge, Downing Street, Cambridge, CB2 3EJ, UK
- ⁹Biology Department, Whitman College, Walla Walla, WA, 99362, USA
- 29 ¹⁰ Biology Department, University of North Carolina at Chapel Hill, NC, 27599, USA
- 30 ¹¹Department of Natural Resource Management, South Dakota State University, SD 57007, USA
- 31 ¹²Department of Biology and Microbiology, South Dakota State University, SD, 57007, USA
- 32 ¹³Department of Ecology and Evolutionary Biology, Tulane University, New Orleans, LA 70118
- 33 ¹⁴Division of Biological Sciences, University of Montana, Missoula, MT, 59812
- 34 ¹⁵Department of Biology, Queen's University, Kingston, Ontario, Canada K7L3N6
- 35 ¹⁶Department of Biology, California Polytechnic State University, San Luis Obispo, CA, 93407,
- 36 USA
- 37 ¹⁷Department of Biological Sciences, Northern Arizona University, Flagstaff, AZ, 86011, USA
- 38 ¹⁸Biological Sciences, California State University, Chico, 95929, USA
- 39 ¹⁹Department of Ecology and Evolutionary Biology, University of California, Santa Cruz, Santa
- 40 Cruz, CA, 95060
- 41 ²⁰Department of Biology and Environmental Science, Auburn University at Montgomery,
- 42 Montgomery, AL, 36124, USA
- 43 ²¹Department of Biology, University of Louisiana, Lafayette, LA, 70503, USA
- 44 ²²Department of Biology, College of Charleston, Charleston, SC, 29424, USA
- 45 ²³Department of Botany, University of Hawai'i, Honolulu, HI, 96822, USA
- 46 ²⁴Environmental Studies Program, University of Colorado Boulder, Boulder, CO 80309, USA

- 47 ²⁵Biology Department, William & Mary, Williamsburg, VA 23187, USA
- 48 ²⁶Ecology & Evolutionary Biology, University of Connecticut, Storrs, CT, 06269, USA
- 49 ²⁷Department of Life and Environmental Sciences, University of California, Merced, CA 95343,
- 50 USA

- 51 ²⁸Department of Plant and Microbial Biology, North Carolina State University, Raleigh, NC,
- 52 27695, USA
- 53 ²⁹Institute of Ecology and Evolution, University of Oregon, Eugene, OR 97403, USA
- 54 ³⁰Department of Genetics, University of Georgia, Athens, GA, 30602, USA
- 55 ³¹Institute of Evolutionary Biology, School of Biological Sciences, Ashworth Laboratories,
- 56 University of Edinburgh, Edinburgh, UK
- 57 ³²Royal Botanic Garden Edinburgh, 20A Inverleith Row, Edinburgh EH3 5LR, UK
- 58 ³³Biological and Environmental Sciences, School of Natural Sciences, University of Stirling,
- 59 Stirling, Scotland, FK9 4LA, UK
- 60 ³⁴Department of Biology, Duke University, Durham, NC, 27708, USA
 - ³⁵Calico Life Sciences LLC, South San Francisco, CA, 94080, USA
 - ³⁶Department of Biology, University of Richmond, Richmond, VA, 23173, USA
 - *Author for correspondence: dlowry@msu.edu

93 The genus *Mimulus* is a well-studied group of plant species, which has for decades allowed 94 researchers to address a wide array of fundamental questions in biology (Wu & al. 2008; 95 Twyford & al. 2015). Linnaeus named the type species of *Mimulus (ringens L.)*, while Darwin 96 (1876) used *Mimulus* (*luteus* L.) to answer key research questions. The incredible phenotypic 97 diversity of this group has made it the focus of ecological and evolutionary study since the mid-98 20th century, initiated by the influential work of Clausen, Keck, and Hiesey as well as their 99 students and collaborators (Clausen & Hiesey 1958; Hiesey & al. 1971, Vickery 1952, 1978). 100 Research has continued on this group of diverse taxa throughout the 20th and into the 21st century 101 (Bradshaw & al. 1995; Schemske & Bradshaw 1999; Wu & al. 2008; Twyford & al. 2015; Yuan 102 2019), and Mimulus guttatus was one of the first non-model plants to be selected for full genome 103 sequencing (Hellsten & al. 2013). *Mimulus* has played a key role in advancing our general 104 understanding of the evolution of pollinator shifts (Bradshaw & Schemske 2003; Cooley & al. 105 2011; Byers & al. 2014), adaptation (Lowry & Willis 2010; Koovers & al. 2015; Peterson & al. 106 2016; Ferris & Willis 2018; Troth & al. 2018), speciation (Ramsey & al. 2003; Wright & al. 107 2013; Sobel & Streisfeld 2015; Zuellig & Sweigart 2018), meiotic drive (Fishman & Saunders 108 2008), polyploidy (Vallejo-Marín 2012; Vallejo-Marín & al. 2015), range limits (Angert 2009; 109 Sexton et al. 2011; Grossenbacher & al. 2014; Sheth & Angert 2014), circadian rhythms 110 (Greenham & al. 2017), genetic recombination (Hellsten & al. 2013), mating systems (Fenster & Ritland 1994; Dudash & Carr 1998; Brandvain & al. 2014) and developmental biology (Moody 111 112 & al. 1999; Baker & al. 2011, 2012; Yuan 2019). This combination of a rich history of study coupled with sustained modern research activity is unparalleled among angiosperms. Across 113 114 many interested parties, the name *Mimulus* therefore takes on tremendous biological significance 115 and is recognizable not only by botanists, but also by zoologists, horticulturalists, naturalists, and 116 members of the biomedical community. Names associated with a taxonomic group of this 117 prominence should have substantial inertia, and disruptive name changes should be avoided. As 118 members of the *Mimulus* community, we advocate retaining the genus name *Mimulus* to describe 119 all monkeyflowers. This is despite recent nomenclature changes that have led to a renaming of 120 most monkeyflower species to other genera.

How did we get here?

121 122

123 124

125

126

127

128

129

130

131

132133

134

135136

137

138

In a recent paper, Barker & al. (2012) proposed splitting the genus *Mimulus* into multiple new genera. This proposed change was based upon a molecular phylogenetic analysis that revealed other small genera, comprising a total of 21 species, were potentially located within the *Mimulus* clade (Figure 1; Beardsley & Olmstead 2002; Beardsley & al. 2004; Beardsley & Barker 2005). The finding that *Mimulus* appears to be a polyphyletic group warranted revision to the genus, as monophyletic groupings are preferred for the designation of genera. Four options were proposed as solutions by Barker & al. (2012): 1) Minimize species name changes by allowing *Mimulus* to remain as a polyphyletic or a biphyletic group; 2) Minimize name changes by grouping all genera into one monophyletic group *Mimulus* L.; 3) Minimize name changes by conserving *Mimulus* L. with a different type species; 4) Divide *Mimulus* into multiple new genera, resulting in many name changes.

Barker & al. (2012) chose to divide *Mimulus* into three major genera (*Mimulus*, *Erythranthe*, and *Diplacus*; Option 4), the solution which required the most name changes (~136 new combinations). They ruled out Option 1, as monophyletic groupings are preferred. They rejected

Option 3, as it would have resulted in name changes to eight widespread *Mimulus* species and would not recognize some genera that the authors designated as distinct. They also stated that they wanted to move forward without waiting for approval of retypification by the next International Botanical Conference in 2017. The justification given for dismissing Option 2 was made based on a desire to conserve the names of a few small Australian genera: "Maximally enlarging *Mimulus* results in the loss of much useful information in the taxonomic hierarchy that recognizes the Australian-centered genera...each of which has apparent apomorphic features that justify treatment at generic rank." Further, it was argued that the Erythranthe and Diplacus clades represented distinct radiations in western North America and that each deserved to be recognized by being elevated to the genus level.

148 149 150

151

152

153

154

155

156

157

158

139

140

141

142

143

144

145

146

147

The nomenclatural suggestions made by Barker & al. (2012) have now been adopted by multiple floras, including the Plants of the World Online, the Oregon Flora Project (Oregon State University), and the Jepson eFlora, and are under review at the Flora of North America. In addition, online resources such as the National Center for Biotechnology Information (NCBI), Global Biodiversity Information Facility (GBIF), UniProt, and iNaturalist now use the names from Barker & al. (2012) in lieu of the older classification. Given the widespread and rapid acceptance of the Barker & al. (2012) circumscription, it may be perplexing as to why so many scientists have continued to use the name *Mimulus*. There are three key reasons why the use of *Mimulus* will likely continue by this group of scientists into the future.

159 160

161

Reason 1: The botanical community needs a stable circumscription for monkeyflowers

162 The primary reason for resisting the adoption of the new nomenclature is that we are reluctant to 163 use different names for the organisms we work on until we are convinced that the nomenclature 164 will be stable for the long-term. Unfortunately, given the limited data to support the name 165 changes, we argue that a conservative position is warranted. In particular, the phylogenetic data 166 available are outdated by modern standards (McKain & al. 2018). Prior to the genomic era, it 167 seemed plausible that sequence data from two genes would be sufficient to approximate the 168 species tree. However, given our modern understanding of the prevalence of gene flow and 169 incomplete lineage sorting (Pease & al. 2016; McKain & al. 2018), it is likely that the species 170 tree for the Phrymaceae will change considerably when more data are added. Prior to revision by 171 Barker & al (2012), the Phrymaceae consisted primarily of the genus *Mimulus*, and several small (in some cases monotypic) genera. Grant (1924) originally separated *Mimulus* into two large sub-172 173 genera based on morphological placentation traits, Synplacus and Schizoplacus (Figure 1), and 174 the taxonomic revisions by Barker & al (2012) elevated these groups to genus level, Erythranthe 175 and *Diplacus* (Figure 1). Our current state of knowledge of the Phrymaceae is based on 176 chloroplast sequence data (trnL/F) and nuclear DNA sequence from the internal and external transcribed spacer nuclear DNA (nrDNA; Beardsley & Olmstead 2002). The chloroplast data 177 178 suggest that the clade containing M. ringens and the Australian Minulus is sister to the clade that 179 includes all other groups, including *Phryma*, *Synplacus*, *Schizoplacus*, and a few other small

180 genera (Figure 1, cpDNA). In contrast, the nrDNA data suggest that *Phryma* is the outgroup to 181

two large clades (Figure 1, nrDNA). One of these clades includes M. ringens, the Australian

182 Mimulus, and subgenus Synplacus. The other clade primarily comprises sub-genus Schizoplacus.

183 When data from the chloroplast gene were combined with the nrDNA data, the resulting 184

topology of the species tree resembled the results from the chloroplast data alone (Beardsley &

185 Olmstead 2002; Beardsley & al. 2004, Beardsley & Barker 2005). This suggests that the 186 chloroplast data were driving the patterns on which Barker's taxonomy was constructed. 187 Unfortunately, trees built from chloroplast data are unreliable because the chloroplast evolves as 188 a single haplotype (McKain & al 2018), frequently spreads to distantly related species by 189 introgression (Rieseberg & Soltis 1991), and often evolves non-neutrally (Wu & Campbell 2007; 190 Bock & al. 2014). Thus, utmost caution is appropriate with regard to the treatment of chloroplast 191 data for phylogenetic questions. We suggest that a modern phylogenetics approach leveraging 192 sequence data from hundreds of nuclear loci and/or an amplicon-based approach incorporating 193 dozens of markers is necessary to gain a better understanding of the species tree topology for the 194 Phrymaceae, as is common in the field (Urive-Convers et al 2016; McKain et al. 2018). The need 195 for more sequence data is illustrated by the comparison of the phylogeny presented in Beardsley 196 & al. (2004) with the modern 41,528-SNP phylogeny from Stankowski & Streisfeld (2015), 197 which shows discordance in the placement of several monkeyflower species.

198 199

200

201

202

203

204

205

206

207

208

209

210

211

212

It is quite possible that new phylogenetic data will completely upend our current understanding of relationships among species in the Phrymaceae. Thus, our position is that no new nomenclatural changes should be adopted until there is a better understanding of the species tree in this group. It has always been our position that it was premature to rename most of the genus *Mimulus* based on two discordant gene phylogenies, as was done by Barker & al. (2012). Our concern is that prematurely switching to new names may cause additional confusion in the literature, particularly if more robust systematic data are consistent with retaining the original name or indicate yet another name change. Thus, we are reluctant to adopt a new circumscription until we are more assured of its stability. We are not alone in our desire for stability, which has been pointed out by others, including Orchard & Maslin (2005): "Taxonomists must recognize that nomenclature is not a plaything of taxonomy, molecular phylogeny, cladistics or any other special interest group. It is a working tool (a filing system) for all biologists, professional and amateur, and for the wider community, and to be meaningful it needs to be as stable as possible. A naming system that continually changes is not a naming system at all and will be discarded or disregarded."

213214215

216

217218

219

220

221

222

223

224

225

226

227228

229

230

Finally, we are concerned about the stability of the current circumscription by Barker & al. (2012) given that it may not have been sufficiently vetted by peer review. The manuscript was published in Phytoneuron, a journal edited by a coauthor on that paper. The editorial policy of Phytoneuron states "Submissions will be reviewed for content and style by the editor, based on his own knowledge and expertise. If deemed appropriate or necessary by the editor, review by other botanical peers will be sought. An indication of the Phytoneuron review process (if beyond the Editor) will appear in the Acknowledgements." The manuscript's acknowledgements in the published paper contain no information about editorial or peer review, other than acknowledging comments "on a late draft" by two colleagues. Subsequent work by Nesom (2014), published in Phytoneuron, is also inconsistent with scientific knowledge of species-level relationships within the section Simiolus of *Mimulus*. For example, Nesom divided annual and perennial populations of M. guttatus into two separate species, Erythranthe guttata and E. microphylla, respectively. Justification for this splitting is directly contradicted by population genetic data, which demonstrate free genetic exchange between annual and perennial populations of M. guttatus (Oneal & al. 2014; Twyford & Friedman 2015). Similarly, Tulig & Nesom (2012) recently elevated several taxa within the *M. aurantiacus* complex to species rank based solely on

- 231 morphological information. Hybrid zones have been well documented for sub-species of M.
- 232 aurantiacus (Thompson 2005), and subsequent work indicates substantial gene flow across these
- points of contact between incompletely isolated taxa (Sobel & Streisfeld 2015, Stankowski et al
- 234 2017). *Mimulus* is arguably one of the most important plant systems in the world for studies of
- speciation, as we know more about how species form in this group than perhaps any other.
- Therefore, the lack of alignment between empirical studies of speciation and taxonomic species
- delimitation seems like an missed opportunity. We thus call for a re-examination and a more
- 238 rigorous review of this systematic treatment in a traditional peer-reviewed journal.

Reason 2: We do not believe that the name changes were necessary

- 241 The splintering of *Mimulus* into multiple genera has primarily been justified based on genera
- 242 with distinct morphological features being nested within the same clade as traditional *Mimulus*
- species. Further, the strongest argument for the new nomenclature is that it has utility in placing
- 244 genus names on groups that have distinct sets of morphological traits. We very much appreciate
- 245 the contributions made by those who have identified taxonomically useful traits, which will
- surely be valuable for future research. However, the desired taxonomic hierarchy for the
- 247 Phrymaceae could be designated with monophyletic sub-genera. The decision to elevate groups
- 248 to the genus level versus the sub-genus level was a subjective nomenclatural decision. The
- 249 differences in placentation cited by Barker et al. (2012) to justify elevating Erythranthe and
- 250 Diplacus to the genus level have long been recognized (Grant 1924) and thus, do not on their
- own necessitate breaking up the genus *Mimulus*.
- We should also point out that there was uncertainty among the taxonomists who made the
- suggested name changes on how to proceed with the nomenclature of this group. For example,
- Nesom (2011) initially renamed the genera *Hemichaena* and *Leucocarpus* to *Mimulus* stating:
- 255 "Hemichaena and Leucocarpus are both justifiably accommodated as groups within the bounds
- of *Mimulus*." And "In case that it proves desirable to maintain the Australian segregate genera,
- and to maintain *Phryma* as a distinct genus, the suggestion by Beardsley and Barker (2005) to
- 258 conserve the name *Mimulus* with a species from within the American lineage is being followed
- 259 (Nesom and N. Fraga, in prep.)." The following year, a reversal of this course of action was
- 260 made with the publication of Barker & al. (2012). The contradictions between Nesom (2011) and
- Barker & al. (2012) clearly illustrate the subjective nature of decisions regarding nomenclature in
- this group and add to our concerns about the stability of its current circumscription.

Reason 3: *Mimulus* is well recognized in the scientific community

- The name changes have already impacted a large number of scientists whose research is focused
- on *Mimulus*. Barker & al. (2012) dismissed the concerns of these scientists and argued that their
- research is focused on "relatively few species." The casual dismissal of the interests of the
- 268 *Mimulus* scientific community by these authors is questionable for four reasons. First, the
- 269 monkeyflower literature encompasses dozens of different *Mimulus* species across the broader
- clade (Vickery 1978; Cooley & al. 2011; Grossenbacher & Whittall 2011; Grossenbacher & al.
- 271 2014; Sobel 2014; Sheth & al. 2014; Sheth & Angert 2014; Chase & al. 2017; Kooyers & al.
- 272 2017; Peng & al. 2017; Li & al. 2018; Medel & al. 2018; Yuan 2019). Second, this argument
- 273 mischaracterizes the size of the research community that studies *Mimulus*. There are now more

274 than 40 labs worldwide that focus their research effort primarily, if not exclusively, on *Mimulus*.

275 Few non-crop genera, beyond Arabidopsis, have this level of research activity. Third, the

276 argument ignores the fact that the instability of nomenclature may cause scientists to lose track

or overlook critical datasets, especially in large genomic (e.g. NCBI) and biodiversity (e.g.

278 GBIF) databases. Finally, this research community identifies primarily by the name *Mimulus* and

279 has spent considerable time over the last two decades building that community under the name

Mimulus. The name Mimulus is widely recognized by our colleagues within the evolution

281 community, by non-plant biologists, and by program officers at the National Science Foundation,

282 Department of Energy, and National Institutes of Health. Instability in the names of these species 283

therefore impedes communication of our discoveries to the broader scientific community and to

funding agencies. For these reasons we have continued to use the name *Mimulus*.

Where do we go from here?

Since we still do not have a good sense of the historical relationships of the taxa in this group of plants, we suggest retaining the name *Mimulus* for all monkeyflowers until more phylogenetic data are available. Ideally, a more robust phylogeny will be developed soon, allowing for a new circumscription to be proposed prior to the International Botanical Congress in 2023. Once that phylogeny has been published, we suggest two routes for retaining the name *Mimulus* across as broad a swath of monkeyflowers as possible based on alternative phylogenetic topologies:

Suggestion 1.

277

280

284

285 286

287 288

289

290

291

292

293

294 295

296

297

298

299

300

301

302

303

304

305 306

307

308

309

310

311

312

313

314

315

316 317

318

319

If *Phryma* is found to be sister to all monkeyflowers, as in the nrDNA tree (Figure 1), we propose that all species within the monophyletic clade containing monkeyflowers be renamed as Mimulus (Option 2 of Barker & al. 2012). Grouping all of the species into one genus, Mimulus, would maximize name stability, which would be especially useful if new data suggest different phylogenetic relationships within the larger clade. While Option 4 of Barker & al. (2012) resulted in the renaming of at least 136 species, Option 2 results in the introduction of only 13 new name combinations. Further, as noted by Nesom (2011) and Barker & al. (2012), most of these species from *Uvedalia*, *Thyridia*, *Elacholoma*, *Hemichaena*, and *Leucocarpus* already have names in Mimulus.

Suggestion 2.

If *Phryma* is found to be nested within the clade containing monkeyflowers, as in the *trnL/F* (Figure 1), we also propose that the entire clade be renamed to *Mimulus*. However, if it is determined that renaming *Phryma* to *Mimulus* is untenable, we suggest that the name *Mimulus* be conserved across a much larger swath of the clade by changing the type species to a species within sub-genus Synplacus (Option 3 of Barker & al. 2012). We propose that Minulus guttatus be designated as the new type species for *Mimulus*, as it is widely studied by scientists and geographically widespread across western North America. Mimulus guttatus occurs within the center of diversity of monkeyflowers, in contrast to *Mimulus ringens*, whose type species status is largely a historical artifact due to the east-to-west direction of exploration of North America by Europeans. The conservation of the name *Mimulus* by changing the type species would likely bring Leucocarpus and Hemichaena into Mimulus. However, M. ringens and the Australian monkeyflowers would likely need to be given a different genus name. Further, a new phylogeny may confirm the results of the nrDNA data and show that M. guttatus and M. ringens are actually

- 320 more closely related to each other than they are to sub-genus *Schizoplacus*. This would
- 321 potentially entail elevating Schizoplacus to the genus level. However, if Schizoplacus is found to
- 322 be sister to Synplacus, we suggest that both sub-genera be named Mimulus following
- 323 retypification. There is precedent for conservative name changes accomplished via designation
- 324 of a new type specimen to maintain a genus name for a larger clade of species, as has been done
- 325 with the genus Acacia (Orchard & Maslin 2005; McNeill & Turland 2011). The justifications for
- 326 conserving Acacia with a new type are very similar to the justifications for conserving Minulus.

- 328 We have provided several reasons above for why we have continued to use *Mimulus* to describe 329 all monkeyflowers. Until modern genomic data can help resolve the considerable uncertainties 330 described above, we will continue to use the name *Mimulus* in publications, presentations, and 331 communication with the general public. In addition, we strongly advocate that Mimulus be used
- 332 in databases and floras until the circumscription of this group is more stable.

333

AUTHOR CONTRIBUTIONS

334 335 336

All authors contributed to the writing of the manuscript.

337 338

LITERATURE CITED

339

340 **Angert, A.L.** 2009. The niche, limits to species' distributions, and spatiotemporal variation in 341 demography across the elevation ranges of two monkeyflowers. P. Natl. Acad. Sci. USA 342 106: 19693–19698.

343 344

Baker, R.L. & Diggle, P.K. 2011. Node-specific branching and heterochronic changes underlie population-level differences in *Mimulus guttatus* (phrymaceae) shoot architecture. *Am. J. Bot.* 98: 1924–1934.

346 347

345

348 Baker, R.L., Hileman, L.C. & Diggle, P.K. 2012. Patterns of shoot architecture in locally 349 adapted populations are linked to intraspecific differences in gene regulation. New Phytol. 196: 350 271-281.

351

352 Barker, W.R., Nesom, G.L., Beardsley, P.M. & Fraga, N.S. 2012. A taxonomic conspectus of 353 Phrymaceae: a narrowed circumscription for Mimulus, new and resurrected genera, and new 354 names and combinations. Phytoneuron 39: 1-60.

355

356 Beardsley, P.M. & Olmstead, R.G. 2002. Redefining Phrymaceae: the placement of *Mimulus*, 357 tribe Mimuleae, and Phryma. Am. J. of Bot. 89: 1093-1102.

358 359

Beardsley, P.M., Schoenig, S.E., Whittall JB & Olmstead RG. 2004. Patterns of evolution in western North American Mimulus (Phrymaceae). Am. J. Bot. 91: 474-489.

360 361 362

Beardsley, P.M. & Barker, W.R. 2005. Patterns of evolution in Australian *Mimulus* and related genera (Phrymaceae~ Scrophulariaceae): a molecular phylogeny using chloroplast and nuclear sequence data. Aust. Syst. Bot. 18: 61-73.

364 365

- Bock, D.G., Andrew, R.L. & Rieseberg L.H. 2014. On the adaptive value of cytoplasmic
- 367 genomes in plants. *Mol. Ecol.* 23: 4899-4911. 368
- 369 Bradshaw, H.D., Wilbert, S.M., Otto, K.G. & Schemske, D.W. 1995. Genetic mapping of
- floral traits associated with reproductive isolation in monkeyflowers (*Mimulus*). *Nature* 376:762-371 765.
- 372

390

394

397

403

- 373 **Bradshaw, H.D. & Schemske D.W.** 2003. Allele substitution at a flower colour locus produces a pollinator shift in monkeyflowers. *Nature* 426: 176-178.
- 375
 376 **Brandvain, Y., Kenney, A.M., Flagel, L., Coop, G. & Sweigart, A.L**. 2014. Speciation and introgression between *Mimulus nasutus* and *Mimulus guttatus*. PLoS Genetics 10: e1004410.
- Byers, K.J.R.P., Vela, J.P., Peng F., Riffell, J.A. & Bradshaw H.D. 2014. Floral volatile
 alleles can contribute to pollinator-mediated reproductive isolation in monkeyflowers (*Mimulus*).
 Plant J. 80: 1031-1042.
- Chase, M.A., Stankowski, S. & Streisfeld MA. 2017. Genomewide variation provides insight into evolutionary relationships in a monkeyflower species complex (*Mimulus* sect. Diplacus).
 Am. J. Bot. 104: 1510-1521.
- 386
 387 Clausen J. & Hiesey W.M. 1958. Experimental studies on the nature of species. IV. Genetic
 388 structure of ecological races. Carnegie Institution of Washington Publication 615. Washington,
 389 DC.
- Cooley A.M., Modliszewski, J.L. Rommel, M. & Willis J.H. 2011. Gene duplication in
 Mimulus underlies parallel floral evolution via independent trans-regulatory changes. *Curr. Biol.* 700-704.
- Darwin, C. 1876. The effects of cross and self fertilisation in the vegetable kingdom. D.
 Appleton and Company, New York, NY, USA.
- Dudash M.R. & Carr D.E. 1998. Genetics underlying inbreeding depression in Mimulus with contrasting mating systems. *Nature* 393: 682.
- Fenster, CB & Ritland, K. 1994. Quantitative genetics of mating system divergence in the yellow monkeyflower species complex. *Heredity* 73: 422-435.
- Fenster, C.B., Diggle, P.K., Barrett, S.C. & Ritland, K. 1995. The genetics of floral development differentiating two species of *Mimulus* (Scrophulariaceae). Heredity 74: 258-266.
- Ferris, K.G. & Willis, J.H. 2018. Differential adaptation to a harsh granite outcrop habitat between sympatric *Mimulus* species. *Evolution* 72: 1225-1241.
- 410 **Fishman, L. & Saunders A.** 2008. Centromere-associated female meiotic drive entails male fitness costs in monkeyflowers. *Science* 322: 1559-1562.

413 **Grant, A.L.** 1924. A monograph of the genus *Mimulus*. *Annals of the Missouri Botanical*

414 Garden 11: 99-388.

415

- 416 Greenham, K., Lou, P., Puzey, J.R., Kumar, G., Arnevik, C., Farid, H., Willis, J.H. &
- 417 **McClung, C.R.** 2017. Geographic variation of plant circadian clock function in natural and
- 418 agricultural settings. J. Biol. Rhythm. 32: 26-34.

419

420 **Grossenbacher, D.L. & Whittall, J.B.** 2011. Increased floral divergence in sympatric monkeyflowers. *Evolution* 65: 2712-2718.

422

- 423 **Grossenbacher, D.L., Veloz, S.D. & Sexton, J.P.** 2014. Niche and range size patterns suggest that speciation begins in small, ecologically diverged populations in North American
- 425 monkeyflowers (*Mimulus* spp.). Evolution 68: 1270-1280.

426

- Hellsten, U., Wright, K.M., Jenkins, J., Shu, S., Yuan, Y., Wessler, S.R., Schmutz, J.,
- Willis, J.H. & Rokhsar, D.S. 2013. Fine-scale variation in meiotic recombination in Mimulus
- inferred from population shotgun sequencing. P. Natl. Acad. Sci. USA 110: 19478-19482.

430

- 431 **Hiesey, W.M., Nobs, M.A. & Björkman, O.** 1971. Experimental studies on the nature of
- species. V. Biosystematics, genetics, and physiological ecology of the *Erythranthe* section of
- 433 *Mimulus*. Carnegie Institute of Washington Publication, 628, 1-213.

434

- 435 Kooyers N.J., Greenlee A.B., Colicchio J.M., Oh M., & Blackman B.K. 2015. Replicate
- altitudinal clines reveal that evolutionary flexibility underlies adaptation to drought stress in
- 437 annual Mimulus guttatus. New Phyt. 206: 152-165.

438

- 439 Kooyers N.J., James B., & Blackman B.K. 2017. Competition drives trait evolution and
- character displacement between *Mimulus* species along an environmental gradient. *Evolution* 71:
- 441 1205–1221.

442

443 **Li, Q., Grossenbacher, D.L. & Angert AL.** 2018. The effect of range overlap on ecological niche divergence depends on spatial scale in monkeyflowers. *Evolution* 72: 2100-2113.

445

- 446 **Lowry, D.B & Willis, J.H.** 2010 A widespread chromosomal inversion polymorphism
- contributes to a major life-history transition, local adaptation, and reproductive isolation. *PLoS*
- 448 Biol. 8: e1000500.

449

- 450 McKain, M.R., Johnson, M.G., Uribe-Convers, S., Eaton, D. & Yang, Y. 2018. Practical
- 451 considerations for plant phylogenomics. *Appl. Plant Sci.* 6: e10.

452

- 453 McNeill, J. & Turland, N.J. 2011. Major changes to the code of nomenclature—Melbourne,
- 454 July 2011. *Taxon* 60: 1495-1497.

- 456 Medel, R., Gonzalez-Browne, C., Salazar, D., Ferrer, P. & Ehrenfeld, M. 2018. The most
- 457 effective pollinator principle applies to new invasive pollinators. *Biol. Lett.* 14: 5.

- 459 **Moody, A., Diggle, P.K. & Steingraeber, D.A.** 1999. Developmental Analysis of the
- 460 Evolutionary Origin of Vegetative Propagules in *Mimulus gemmiparus* (Scrophulariaceae). *Am.*
- 461 *J. Bot.* 86: 1512–1522.

462

- Nesom, G.L. 2011. Recognition and synopsis of *Mimulus* sect. *Tropanthus* and sect.
- 464 Leucocarpus (Phrymaceae). Phytoneuron 28: 1–8.

465

- Nesom, G.L. 2014. Updated classification and hypothetical phylogeny of *Erythranthe* sect.
- 467 Simiola (Phrymaceae). *Phytoneuron* 81: 1–6.

468

- Oneal, E., Lowry, D.B., Wright, K.M., Zhu, Z. & Willis, J.H. 2014. Divergent population
- 470 structure and climate associations of a chromosomal inversion polymorphism across the M
- imulus guttatus species complex. *Mol. Ecol.* 23: 2844-2860.

472

- 473 **Orchard, A.E., Maslin, B.R.** 2005. The case for conserving *Acacia* with a new type. *Taxon*. 54:
- 474 509-512.

475

- Pease, J.B., Haak, D.C., Hahn, M.W., Moyle, L.C. 2016. Phylogenomics reveals three sources
- of adaptive variation during a rapid radiation. *PLoS Biol.* 14: e1002379.

478

- 479 **Peng F., Byers K.J., & Bradshaw H.D.** 2017. Less is more: Independent loss-of-function
- 480 OCIMENE SYNTHASE alleles parallel pollination syndrome diversification in monkeyflowers
- 481 (Mimulus). Am. J. Bot. 104: 1055-1059.

482

- 483 **Peterson, M.L., Kay, K.M. & Angert, A.L.** 2016. The scale of local adaptation in Mimulus
- guttatus: comparing life history races, ecotypes, and populations. *New Phytol.* 211: 345-356.

485

- 486 Ramsey, J., Bradshaw, H.D., Schemske, D.W. 2003. Components of reproductive isolation
- between the monkeyflowers *Mimulus lewisii* and *M. cardinalis* (Phrymaceae). *Evolution* 57:
- 488 1520-1534.

489

- 490 **Rieseberg, L.H. & Soltis, D.E.** 1991. Phylogenetic consequences of cytoplasmic gene flow in
- 491 plants. Evol. Trend. Plant. 5: 65-84.

492

- 493 **Schemske, D.W. & Bradshaw, H.D.** 1999. Pollinator preference and the evolution of floral
- 494 traits in monkeyflowers (Mimulus). P. Natl. Acad. Sci. USA 96: 11910-11915.

495

- 496 **Sexton, J.P., Strauss, S.Y., & Rice, K.J.** 2011. Gene flow increases fitness at the warm edge of
- 497 a species' range. P. Natl. Acad. Sci. USA 108:11704-11709.

498

- 499 **Sheth, S.N. & Angert, A.L.** 2014. The evolution of environmental tolerance and range size: a
- comparison of geographically restricted and widespread *Mimulus*. *Evolution*. 68: 2917-2931.

- 502 Sheth, S.N, Jiménez, I., & Angert, A.L. 2014. Identifying the paths leading to variation in
- 503 geographical range size in western North American monkeyflowers. J. Biogeogr. 41: 2344-2356.

505 **Sobel, J.M.** 2014 Ecogeographic isolation and speciation in the genus *Mimulus*. *Am. Nat.* 184: 506 565-579.

507

508 Sobel, J.M. & Streisfeld, M.A. 2015. Strong premating reproductive isolation drives incipient 509 speciation in Mimulus aurantiacus. Evolution 69: 447-461.

510

511 Stankowski, S., & Streisfeld, M.A. 2015. Introgressive hybridization facilitates adaptive 512 divergence in a recent radiation of monkeyflowers. *Proceedings of the Royal Society B:* Biological Sciences 282: 20151666.

513

514

515 Stankowski, S., Sobel, J.M., Streisfeld, M.A. 2017. Geographic cline analysis as a tool for 516 studying genome-wide variation: a case study of pollinator-mediated divergence in a 517 monkeyflower. Mol. Ecol. 26: 107-122.

518

519 **Thompson, D.M.** 2005. Systematics of *Mimulus* subgenus *Schizoplacus* (Scrophulariaceae). 520 Syst. Bot. 75: 1-213.

521

522 Tulig, M.C., & Nesom, G.L. 2012. Taxonomic overview of Diplacus sect. Diplacus 523 (Phrymaceae). Phytoneuron 45: 1-20.

524

525 Troth, A., Puzey, J.R., im, R.S., Willis, J.H. & Kelly, J.K. 2018. Selective trade-offs maintain 526 alleles underpinning complex trait variation in plants. Science 361: 475-478.

527 528

Twyford, A.D. & Friedman J. 2015 Adaptive divergence in the monkeyflower *Mimulus* guttatus is maintained by a chromosomal inversion. Evolution 69: 1476-1486.

529 530

531 Twyford, A.D., Streisfeld, M.A., Lowry, D.B. & Friedman, J. 2015 Genomic studies on the 532 nature of species: adaptation and speciation in Mimulus. Mol. Ecol. 24: 2601-2609.

533

534 Uribe-Convers, S., Settles, M.L., & Tank, D.C. 2016. A phylogenomic approach based on 535 PCR target enrichment and high throughput sequencing: Resolving the diversity within the South 536 American species of Bartsia L. (Orobanchaceae). PLoS One 11: e0148203.

537

538 Vallejo-Marín, M. 2012. Mimulus peregrinus (Phrymaceae): A new British allopolyploid 539 species. Phytokeys 14: 1-14.

540

541 Vallejo-Marín, M., Buggs, R. J., Cooley, A. M., & Puzey, J. R. 2015. Speciation by genome 542 duplication: Repeated origins and genomic composition of the recently formed allopolyploid 543 species Mimulus peregrinus. Evolution 69: 1487-1500.

544

545 Vickery, R.K. 1952. A study of the genetic relationships in a sample of the *Mimulus guttatus* 546 complex. PhD Dissertation Dept. of Biological Sciences, Stanford University.

547

548 Vickery, R.K. 1978. Case studies in the evolution of species complexes in *Mimulus*. Evol. Biol. 549 11: 405-507.

Wright, K.M., Lloyd, D., Lowry, D.B., Macnair, M.R. & Willis JH. 2013. Indirect evolution of hybrid lethality due to linkage with selected locus in Mimulus guttatus. PLoS Biol. 11, e1001497. Wu, C.A. & Campbell, D.R. 2007. Leaf physiology reflects environmental differences and cytoplasmic background in *Ipomopsis* (Polemoniaceae) hybrids. Am. J. Bot. 94: 1804-1812. Wu, C.A., Lowry, D.B., Cooley, A.M., Wright, K.M., Lee, Y.W. & Willis, J.H. 2008. Mimulus is an emerging model system for the integration of ecological and genomic studies. Heredity 100: 220-230. Yuan, Y.W. 2019. Monkeyflowers (Mimulus): new model for plant developmental genetics and evo-devo. New Phytol. 222: 694-700. Zuellig M.P. & Sweigart A.L. 2018. A two-locus hybrid incompatibility is widespread, polymorphic, and active in natural populations of *Mimulus*. Evolution 72: 2394-2405.

FIGURE LEGENDS

596 597 598

599

600 601

602

603

604

605

606

607

608

609

610

611

612

Figure 1. Phylogenetic hypotheses in the monkeyflowers (redrawn from data presented in Beardsley and Olmstead 2002 and Beardsley & al 2004). The type specimen for the genus Mimulus is the eastern North American species M. ringens, which appears to be sister to a radiation of Australian taxa. Grant (1924) separated Minulus into two sub-genera based on morphological traits, Synplacus and Schizoplacus. These highly diverse groups are further divided into cohesive morphological sections that contain several well-studied ecological model systems, such as M. guttatus, M. lewisii, and M. aurantiacus. Phylogenies based on DNA sequences indicate that several very small genera are nested within the diversity present in Mimulus: e.g. Leucocarpus, Berendtiella, Hemichaena, and Phryma. However, phylogenetic hypotheses are based on only a small number of chloroplast (trnL/F) and nuclear loci (ITS/ETS), and substantial uncertainty exists at levels relevant to recent taxonomic revisions. For example, Phryma (dashed) is placed sister to the entire group according to the nuclear loci (nrDNA), and nested within *Mimulus* for the chloroplast locus (cpDNA). Further, the placement of *M. ringens* and related Australian species is uncertain (bold), with nrDNA indicating them to be sister to subgenus Synplacus, and cpDNA placing them sister to the entire group. Bold and dashed branches are used to highlight discordances between the nrDNA and cpDNA phylogenies.