

1 *Invited Review for Current Biology: 3000-4000 words, at least 3 figures, at least 50 refs*

2 **Mutualisms and (a)symmetry in plant-pollinator interactions**

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16

17 **Summary**

18 The majority of flowering plants relies on animal pollinators for sexual reproduction and
19 many animal pollinators rely on floral resources. However, interests of plants and pollinators
20 are often not the same, resulting in an asymmetric relationship that ranges from mutualistic to
21 parasitic interactions. Our understanding of the processes that underlie this asymmetry
22 remains fragmentary. In this review, we bring together evidence from evolutionary biology,
23 plant chemistry, biomechanics, sensory ecology and behaviour to illustrate that the degree of
24 symmetry often depends on the perspective taken. We also highlight variation in (a)symmetry
25 within and between plant and pollinator species as well as between (geographic) locations.
26 Through taking different perspectives from the plant and pollinator side we provide new
27 ground for studies on the maintenance and evolution of animal pollination and on the
28 (a)symmetry in plant-pollinator interactions.

29

30 **Keywords:** chemical ecology, biophysics, pollination, reproduction, evolution

31 **Introduction**

32 The fascinating interaction between plants and pollinators is sometimes referred to as a
33 mutual partnership [e.g. 1], but the interaction is driven by the different and often conflicting
34 interests of plants and pollinators. Plants need reliable pollen dispersal and receipt at minimal
35 costs, whereas pollinators seek floral rewards that can be harvested as fast and efficient as
36 possible. The resulting interaction is sometimes referred to as (balanced) mutual exploitation
37 [sensu 2].

38 Plants and pollinators have evolved traits that influence the balance of this mutual
39 exploitation to their advantage [3–5] (Figure 1). Plants essentially face two connected
40 challenges: (1) How to optimise the (amount of) reward offered to actual pollinators? (2)
41 How to prevent access to floral resources by flower visitors that collect rewards but do not
42 pollinate? To paraphrase Danforth and colleagues [3]: pollinators are like overly demanding
43 lovers: they are great to have around, but if left without boundaries, they will take over your
44 life and ruin it. Many plant adaptations restrict access to floral rewards in a variety of ways,
45 ranging from morphological to mechanical or chemical boundaries. Conversely, pollinators
46 evolved strategies to increase exploitation of floral resources to their advantage, such as
47 morphological, behavioural and/or physiological adaptations.

48 The suite of (co-evolved) adaptations mediates a tension between plants and
49 pollinators, which results in different types of interactions. The (relative) benefit for plants
50 versus pollinators ranges from solely beneficial for flower visitors (e.g. reward robbing) via
51 more balanced interactions to solely beneficial for plants (e.g. pollination by deception;
52 Figure 1). Understanding the different aspects of the tension between plants and pollinators
53 calls for a cross-disciplinary view, because the traits involved span across different scientific
54 domains.

55 In this review, we discuss classic and recent work on the chemical, mechanical and
56 sensory aspects of plant-pollinator interactions, and how work from various disciplines can
57 help understanding the transition from mutualism to antagonism. We also argue that the
58 degree of symmetry in the interaction between plants and pollinators is more variable than
59 often appreciated, and depends on the perspective taken, species and/or geographic location.

60

61 **Plant reward chemistry and pollinator dietary preferences**

62 Many pollinators, in particular bees, depend on pollen, nectar and/or floral oils as major or
63 sole food sources. Except for pollen, which primary function is to fertilise ovules, the only
64 function of other types of rewards is to reward pollinators. Their chemical/nutritional

65 composition (mainly protein, lipids, sugars and micro-nutrients) could therefore be adjusted
66 to the needs of a specific pollinator group and be under selective pressure by pollinators
67 (Figure 2). However, with few exceptions (e.g. specialized oligolectic bees restricted to few
68 plant species/genera), the nutritional composition of floral rewards often does not meet the
69 specific dietary preferences of potential pollinators. For instance, bumblebees (*Bombus*
70 *terrestris* and *B. impatiens*) were found to preferentially collect pollen with a protein to lipid
71 (P:L) ratio of between 5:1 and 10:1 [6], but that ratio is found in only a minority of species,
72 viz. 33 out of 86 (38%) plant species [7]. Similarly, the majority of the 82 plant species
73 chemically analysed by Vaudo and colleagues [8] fell within P:L ratios lower than 3:1. By
74 the same token, honeybee (*Apis mellifera*) workers prefer nectar with a protein to
75 carbohydrate (P:C) ratio of 1:250 [9], which was found in only three out of 304 nectar
76 samples (<1%) from overall 34 plant species (C Venjakob, AM Klein and SD Leonhardt,
77 unpublished data). Recent evidence suggests that preferences for specific nutrient contents
78 and ratios are linked to reproductive fitness (for *B. terrestris* see e.g. [10] and [11]). If
79 (foraging) preferences generally mirror nutritional requirements and are linked to pollinator
80 fitness still needs to be elucidated.

81 In addition to nutrients, pollen and nectar often contain significant amounts of
82 species-specific plant secondary metabolites (PSMs, [12]), including alkaloids, fatty acid
83 derivatives, different glycosides, flavonoids and terpenoids). Many of these compounds are
84 toxic if consumed at high doses [e.g. toxins in *Aconitum* nectar; 11] (Figure 2). PSMs
85 decrease reward palatability [14,15] and often require specific physiological adaptations,
86 such as specific enzymes (e.g. of the cytochrome family), for detoxification [e.g. 16]. The
87 amount and composition of PSMs can differ between floral rewards, i.e. pollen and nectar, of
88 the same plant [12]. These qualitative and quantitative differences can increase attractiveness
89 of one resource while simultaneously decreasing attractiveness of the other. For example, in
90 two *Dipsacus* species the PSM dipsacus saponin occurs in pollen but not nectar, which
91 renders pollen unattractive to some bumblebee species, resulting in less grooming and thus
92 increased pollen deposition on stigmas [17]. Moreover, PSMs can be toxic or repellent to
93 some flower visitors (or herbivores) and attractive or beneficial to others [reviewed by 18].
94 PSMs in floral rewards can therefore be a filter mechanism and restrict the spectrum of
95 flower visitors to actual pollinators, which generally increases pollination success and
96 pollinator benefits [18,19]. The question why not all plants use PSMs to restrict visitor
97 spectra to preferred pollinators remains to be answered.

98 Various physiological strategies help pollinators to deal with nutritionally imbalanced
99 pollen and nectar diets and to mitigate negative effects of PSMs (Figure 3). For example,
100 eusocial honeybees (*Apis mellifera*) and some species of stingless bees (Apidae: Meliponini)
101 feed their offspring with glandular secretions rather than with pure pollen and nectar
102 [reviewed by 20,21]. Such secretions match nutritional requirements and protect bee larvae
103 from PSMs [22]. Some primitively eusocial bumblebees as well as some solitary bees also
104 add nutritive salivary secretions to nectar and/or pollen prior to feeding larvae (as e.g. shown
105 for *B. terrestris*, [23], and *Anthophora*, [24]). Other (solitary) species are able to digest toxic
106 compounds with the help of specialised digestive systems or specific gut microbiota [25],
107 which allows them to tolerate toxic PSMs. Interestingly, the evolution of tolerance for
108 specific PSMs appears to independent of dietary specialization (i.e. oligolecty vs. polylecty)
109 and shows no phylogenetic pattern [18].

110 Pollinators can also learn to avoid harmful reward compounds and adjust their
111 foraging behaviour through complementary foraging (Figure 3). Diversified reward
112 collection (i.e. allocating and mixing floral resources from different plant species) benefits
113 pollinators through increasing overall resource quantity/quality, diluting toxic compounds or
114 decrease costs associated with handling time (e.g. through mixing resources from
115 morphologically similar flowers) [26–29]. Such mixed foraging likely requires cognitive
116 flexibility and the ability to rapidly assess reward quality (e.g. through prioritized perception,
117 [10]) in order to both readily switch to new floral sources and quickly learn to handle flowers
118 and access rewards (Figure 3). Whether cognitive and behavioural flexibility is associated
119 with an increased tendency of resource mixing has, however, not yet been investigated.

120 Pollinators can also adjust their way of feeding to nectar sugar concentrations in
121 various ways, as shown by fluid dynamic models combined with pollinator experiments. For
122 example, bees, which consume nectar via dipping their tongue, typically feed on nectar with
123 50-60% sugar concentration, whereas butterflies and birds, which feed via suction, typically
124 feed on nectar with 35% sugar concentration, likely because nectar viscosity quickly
125 decreases uptake via suction [30]. Intriguingly, honeybees can switch between suction and
126 lapping, depending on the sugar content and viscosity, which increases their foraging
127 efficiency [31]. Eusocial pollinators like honeybees and bumblebees store collected nectar in
128 nests, which requires foragers to regurgitate nectar from crops. Recent work suggests that
129 offloading complicates maximum energy transfer rates [32], because the offloading process
130 also has implications for efficiency and energy-transfer-rate.

131

132 **Morphological gate keeping of floral rewards, and how to access them**

133 Natural selection favours plant traits that (1) increase the pollen transfer efficiency of
134 pollinators, and (2) exclude antagonists such as herbivores and floral robbers (Figure 2).
135 However, total pollen export to multiple recipient stigmas does not increase linearly with the
136 amount of pollen placed on the bee's body, but rather follows a pattern of diminishing return
137 [33]. Thus, gradual placing of pollen on the right pollinator and body part is often key to
138 outcrossing success [33,34] and decreases the influence of stochastic effects (e.g. death of
139 pollinators before pollen transfer).

140 Plants evolved a range of specialised biomechanical adaptations to regulate access to
141 rewards and optimise pollen transfer (Figure 2), including keel flowers [35], lever-
142 mechanism flowers [36,37], trigger flowers [38,39], explosive pollen release [40,41],
143 heteranthery [42], and buzz-pollinated flowers [43]. For example, keel flowers present in
144 many Fabaceae and Polygalaceae require floral visitors to exert a significant amount of force
145 to access nectar and pollen rewards [37,44]. As the bee pushes its body to access nectar, the
146 keel is pushed downwards, exposing pollen that is deposited on the bee's body. Keel
147 morphology acts effectively as a pollinator filter, because it excludes floral visitors that do
148 not have the physical attributes (e.g., butterflies) or strength (small bees, flies) to push open
149 these flowers [44]. The keel morphology may also contribute to regulating pollen dispensing
150 [45] and pollen placement on the visitor's body [46]. These three roles (pollinator filtering,
151 pollen dispensing, and improved pollen placement) are not mutually exclusive and probably
152 jointly explain the evolution of the keel and other specialised floral morphologies.

153 The anther-lever mechanisms, such as are present in most of *Salvia* (Lamiaceae)
154 species, provide another example of how flower biomechanics filter pollinators by requiring a
155 specific amount of strength to access rewards (Figure 2). The stamens form a physical barrier
156 to the nectar, and the floral visitor (e.g. bees, moths or hummingbirds) needs to push through,
157 bringing the anthers into contact with the dorsal side of the floral visitor [47]. Medium and
158 large-sized bees can exert forces of 20-100 mN when visiting *Salvia*, which is enough to
159 trigger the lever mechanism [36,44]. Biomechanical adaptations aimed at excluding some
160 visitors thus seem most effective against insects other than bees, such as flies, butterflies and
161 moths.

162 Buzz pollination further illustrates how floral visitors have driven the evolution of the
163 biomechanical properties of flowers. Buzz-pollinated flowers often display anthers that open
164 through small gaps or pores (poricidal anthers) instead of laterally dehiscent as most flowers
165 do. During buzz pollination, bees produce vibrations to remove pollen from flowers, which

166 are also usually nectarless [48]. Poricidal anthers thus act as filters excluding or discouraging
167 visits by some animals (*e.g.*, flies, butterflies, non-buzzing bees) (Figure 2). In addition,
168 poricidal anthers may also improve pollination efficiency by acting as pollen dispensers in
169 which pollen release by individual buzzing bees is physically restricted [49]. Gradual pollen
170 dispensing allows distributing pollen loads across more floral visitors, which can increase
171 reproductive success when visitation rates are high and pollen delivery uncertain [33,50].
172 In turn, pollinators have evolved different behavioural characteristics that enable them to
173 most efficiently exploit floral resources (Figure 3). For example, more than 50% of bee
174 species, including bumblebees, carpenter bees and euglossine bees, are able to use their
175 (flight) muscles to produce vibrations on flowers, *i.e.* buzz flowers, and so collect pollen
176 from buzz-pollinated flowers [51]. The evolution of buzz pollination involves the co-option
177 of a pre-existing behaviour by bees (buzzing) [52] to increase the rate of pollen removal from
178 flowers, and the co-evolution of increasingly restrictive floral morphologies, such as poricidal
179 anthers, that restrict and regulate pollen removal [53]. The vibrations produced vary widely in
180 frequency, amplitude and duration, properties which determine the amount of pollen released
181 from buzz-pollinated flowers [43,53,54]. They presumably are a function of the bees'
182 morphological, physiological and neurological characteristics, but little is known about what
183 exact characteristics determine the vibrations bees produce and how this varies between
184 species [55].

185 Pollinator body size also plays an important role in several other interaction contexts.
186 For example, bees with larger bodies can save energy owing to a higher surface to volume
187 ratio, and so fly longer distances [56]. Large bees can thus cover large foraging areas, which
188 may support floral resource mixing through collecting rewards from a greater diversity of
189 flowers. Larger bodies or mandibles may further facilitate robbing in bees, allowing species
190 to circumvent morphological features of plants (*i.e.* elongated tubes, poricidal anthers, keel
191 flowers, anther-lever) aimed at restricting access to specific pollinators. Conversely, small
192 bees may bypass physical barriers and so rob nectar while failing to trigger the lever
193 mechanism thus removing the reward without pollination (Figure 3). Notably, robbing is a
194 facultative behaviour, as most robbers also forage regularly on flowers [57] thereby providing
195 the actual pollination service. It hence seems unlikely that pollinators evolved specific traits
196 to facilitate robbing, but rather use pre-existing ones, such as large/strong maxillae or small
197 elongated bodies.

198

199 **Sensory interplay between plants and pollinators**

200 Flowers create signals via colour, odour and/or shape, and pollinators perceive these signals
201 via visual, olfactory and/or tactile mechanisms, which involve various biophysical processes.
202 To be detected by pollinators, flowers have to stand out from the environment; for example,
203 by producing colours that contrast with the background [58] and/or via emitting attractive
204 scents [59]. Further, to promote repeated visitation, flowers should be memorably rewarding,
205 which can be achieved by generating a stimulus that the pollinator will associate with a
206 reward.

207 To attract pollinators – ideally while simultaneously being cryptic to antagonists –
208 plants often combine different types of signals. Sensory signals can thus be used to
209 selectively inform pollinators that perceive specific signals, while excluding antagonists
210 (Figure 2). For example, most pollinating insects have very poor visual sensitivity in the red
211 part of the light, which creates a “private niche” in colour space for birds, which do see red
212 colours [60]. However, colour can only work as pollinator-selective signal when pollinators
213 and non-pollinators have different visual systems. In the case of bees, of which there are both
214 pollinators and robbers, colour vision is highly similar among species [61,62], thus
215 preventing colour as sole filter. In these cases, scent, shape or patterning can aid deterring or
216 at least reducing conspicuousness to antagonists. The morphologically similar flowers of two
217 related *Gomphocarpus* species (Apocynaceae), for example, attract pollinators (bees versus
218 spicid wasps) mostly by emitting different scents [63]. Floral scent also seems to be the
219 principal mechanism that determines pollinator specificity in orchids pollinated by euglossine
220 male bees, who collect the chemical compounds and mix them to produce perfumes that
221 attract females [64]. Moreover, a meta-analysis that compared the attractiveness of floral
222 scents to pollinators and antagonists suggested that scents can serve the dual function of both
223 attracting pollinators and deterring antagonists [65].

224 Some plant species alter floral signals, for example after pollination to deter
225 pollinators (and florivores) from already pollinated flowers (Figure 2). Flowers that lose
226 colour are less attractive to pollinators, but still enhance long-distance attraction of the
227 inflorescence as a whole [66], because the unpigmented, pale flowers generate a bright signal
228 that will be visible from long-distances [58]. Flower visibility can also be reduced
229 temporarily by closing inflorescences after pollination, which reduces conspicuousness to
230 antagonists [67], due to changes in inflorescence display size/shape and a less colourful lower
231 side of the florets [68,69]. Scent emission is more plastic than colour, and (diel) patterns in
232 scent emission frequently match anthesis and nectar availability, particularly for nocturnal
233 pollinated flowers [70,71]. In *Silene latifolia*, scent emission decreases more rapidly in

234 pollinated than unpollinated flowers [72], likely resulting in decreased visitation by
235 pollinators and antagonists.

236 At the extreme, floral signals can also be used to deceive pollinators (Figure 2).
237 Deception of pollinators, where flowers mimic food, mating or oviposition sites, has evolved
238 in at least 7500 species (6%) across 32 animal-pollinated angiosperm families [73].
239 Intriguingly, deception of pollinators occurs almost exclusively in insect-pollinated plants,
240 but whether this is due to insects being more easily cheated upon or because the vast majority
241 of animal pollinators are insects is unknown. Deception often occurs because flowers
242 generate signals that capitalise on innate behavioural responses of pollinators [74], which can
243 have evolved outside the context of plant-pollinator interactions [75]. Innate colour
244 preferences, for example, occur in many groups of flower-visiting insects, including flies,
245 butterflies and bees [58,76,77], and have been thought to be important as a deceptive cue in,
246 for example, orchids [78]. Combinations of fine-tuned visual, chemical and tactile signals can
247 yield floral phenotypes that are deceptive to the point that pollinators choose the deceptive
248 flower over the genuine object the flower mimics. For example, sexually deceptive *Ophrys*
249 flowers are preferred by male wasps over a female conspecific [79]. In addition to colour and
250 scent, some plant species evolved flower thermogenesis (heat production), e.g. they mimic a
251 suitable egg-laying substrate, such as dung or carrion, and so co-opt innate cues used by
252 insects to find oviposition sites [75]. However, visiting a warm, albeit rewardless, flower
253 need not always be bad, because heat in itself can be a reward to insect pollinators,
254 particularly when ambient temperatures are low [80].

255 In response to deception, pollinators can learn to avoid deceptive flowers after one or
256 a few visits [81], which impedes future outcrossing (Figure 3). Indeed, aversive learning of
257 inflorescence hue [82], colour patterns [83] and odour [84] occurs across insects that
258 pollinate sexually deceptive flowers, and this is expected to lead to negative frequency-
259 dependent selection of flower morphs. The situation becomes trickier when only some
260 flowers on a plant are deceptive, because these few flowers may result in the entire individual
261 suffering from reduced pollination. For example, pollinating bumblebees rapidly learn to
262 detect and avoid rewardless (female) *Begonia odorata* flowers, which comes at the expense
263 of visiting real (male) flowers that are highly similar in terms of visual signals [85]. Overall,
264 bumblebees nevertheless benefit from their learning ability, because the time saved by
265 avoiding rewardless flowers outweighs the effect of missing out on genuine flowers [85].
266 There are situations, however, where avoiding deceptive flowers becomes detrimental for the
267 pollinators' own fitness. For example, male wasps learn to avoid areas with sexually

268 deceptive *Chiloglottis trapeziformis* flowers [86]. As a consequence, they also avoid female
269 wasps occurring in the same areas as the plants, which reduces mating events [86].

270 In the absence of extreme cases such as (sexual) deception, pollinators can also learn
271 to use particular floral cues as indicators for reward quantity and/or quality (Figure 3). For
272 example, bumblebees learn to associate floral scent compounds with the amount of reward in
273 *Brassica rapa* and *Penstemon digitalis* [87,88]. In this case, distinct chemical cues are good
274 candidates to honestly signal reward quantity [89]; however, flowers typically provide a
275 plethora of (chemical) signals. To make sense of this signal complexity, pollinators may
276 either simultaneously process multimodal information [90] or prioritise perception of those
277 cues that are linked to compounds that are most influential (e.g. affect reproductive fitness),
278 such as specific nutrients or toxic compounds, while ignoring others [10]. Taken together, the
279 sensory interplay between plants and pollinators represents an impressive array of partly
280 highly species-specific fine-tuning that involves a variety of biochemical and neuronal
281 adaptations.

282

283 **Variation in time and space, and phenotypic mosaics**

284 Spatial or temporal variation in plant or pollinator phenotype, abundance or community can
285 create mosaic-like patterns in the degree of mutualism [91]. In addition to interspecific
286 variation, phenotypic variation can occur between populations, individuals and even within
287 an individual, as is the case for plants bearing multiple flowers at a given moment. A neat
288 example of geographic variation in the degree of plant-pollinator mutualism is that of
289 *Lithophragma parviflorum* and its pollinator, *Greya pollitella* moths [4,92]. *Greya pollitella*
290 are effective pollinators, though their larvae consume a small number of developing seeds.
291 When *Greya* moths are the sole pollinators, their pollination service outweighs the
292 antagonistic effects caused by the larvae. However, in populations with many bombyliid flies,
293 which are equally efficient pollinators but do not parasitise developing seed, the interaction
294 with *Greya* moths becomes antagonistic [4].

295 Floral reward quantity and quality, as well as pollination service are often dynamic in
296 both time and space. For plants, such variation in reward quality and availability occurs at
297 various levels [88,93,94]. Predictable within-plant variation in reward availability can modify
298 the degree of geitonogamy and thereby increase plant reproductive success. In vertical
299 inflorescences, for example, a decreasing amount of nectar from lower to upper flowers, can
300 encourage pollinators to leave a plant and go to another individual when facing a decreasing
301 amount of reward per flower [95]. At the community level, variation in reward availability

302 may result in increased pollen dispersal, because pollinators tend to fly longer distances after
303 encountering rewardless flowers [96].

304 Regional and temporal differences in pollinator community can in turn impose
305 varying selective pressures on floral traits. Geographic covariation of floral and pollinator
306 traits has been reported for numerous traits, such as flower morphology and nectar properties
307 versus bird beak morphology [97], optical properties and colour vision [98,99] and scent and
308 pollinator community [100]. All else being equal, spatially heterogeneous selection on any
309 trait results in a non-homogeneous trait landscape. For example, in the spring ephemeral
310 *Claytonia virginica*, populations of plants that are pollinated by pollen-collecting bees
311 produced more pollen per flower and showed more staggered pollen release than populations
312 pollinated by nectar-foraging bee-flies [101]. Such examples highlight local adaptation of
313 individual plants to the present pollinator fauna, resulting in variable degrees of (a)symmetry
314 even within the same plant species.

315 Studies on variation in floral rewards mostly focused on nectar properties, and how
316 observed variation relates to diel patterns, pollinator activity, abiotic effects and/or flower age
317 [95]. We lack knowledge on rewards other than nectar, how they vary in quantity (e.g.
318 amount of heat as a reward) and quality (e.g. pollen chemical profile), and to what degree
319 their value to pollinators is context-dependent.

320

321 **Conclusions**

322 The interaction between plants and pollinators is characterised by a tension arising from
323 conflicting interests of both parties. Evolutionary adaptations in plants and pollinators can
324 lead either to a balance between partners resulting in a symmetric relationship, or to an
325 imbalanced, asymmetric relationship. How symmetric or asymmetric interactions between
326 plants and pollinators really are is hard to determine, as information on pollination efficacy
327 and/or actual nutritional requirements and tolerance of pollinators is lacking.

328 Relative benefits of plants and pollinators may or may not be balanced in the long
329 term, but the frequently observed asymmetries suggest an evolutionary arms race.

330 Understanding the sequence of evolutionary events will be useful to understand that putative
331 arms race. For example, the colours and scents of flowers probably evolved long after the
332 pollinator sensory systems that perceive them, and not vice versa [102,103], but how does
333 this apply to other traits, such as reward quality versus pollinator dietary requirements, or
334 restricted floral morphologies and the way pollinators circumvent these? In specialised
335 interactions where plants and pollinators strongly depend on each other, pollinator and plant

336 adaptations probably evolved synchronously [e.g. bees' ability to sonicate and poricidal
337 anthers; 100]. Other taxa that were not originally part of the interaction may have evolved
338 similar traits at a later stage, however. Capitalising on (repeated) switches between
339 pollination systems [34,104,105], pollinator behaviour (e.g. buzz pollination; [52]) or floral
340 phenotypes, is a powerful way to explore the evolutionary trajectory of adaptations and how
341 they may have led to the extant interaction.

342 The degree of asymmetry often varies in time and space, which complicates matters
343 further. Determining the factors that underlie inter- and intraspecific variation in the type,
344 quantity, quality and accessibility of rewards across time and space, and how pollinators
345 respond to such variation is central to understanding the degree of (a)symmetry in different
346 plant-pollinator interactions. This is particular true for rewards that are dynamic and context-
347 dependent (e.g. heat or mating opportunities).

348

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355

356 **References**

- 357 1. Barth, F.G. (1991). *Insects and flowers. The biology of a partnership.* (Princeton:
358 Princeton University Press).
- 359 2. Westerkamp, C.H. (1996). Pollen in bee-flower relations some considerations on
360 melittophily. *Bot. Acta* 109, 325–332.
- 361 3. Danforth, B.N., Minckley, R.L., Neff, J.L., and Fawcett, F. (2019). Bees and plants:
362 love story, arms race, or something in between? In *The solitary bees Biology,*
363 *Evolution, Conservation.* (Princeton: Princeton University Press), pp. 289–317.
- 364 4. Anderson, B. (2015). Coevolution in mutualisms. In *Oxford University Press Oxford,*
365 *UK, J. L. Bronstein, ed. (Oxford University Press).*
- 366 5. Bronstein, J.L. (2001). The exploitation of mutualisms. *Ecol. Lett.* 4, 277–287.
- 367 6. Vaudo, A.D., Patch, H.M., Mortensen, D.A., Tooker, J.F., and Grozinger, C.M.
368 (2016). Macronutrient ratios in pollen shape bumble bee (*Bombus impatiens*) foraging
369 strategies and floral preferences. *Proc. Natl. Acad. Sci. U. S. A.* 113, E4035-42.

- 370 7. Ruedenauer, F.A., Spaethe, J., van der Kooi, C.J., and Leonhardt, S.D. (2019).
371 Pollinator or pedigree: which factors determine the evolution of pollen nutrients?
372 *Oecologia* 191, 349–358.
- 373 8. Vaudo, A.D., Tooker, J.F., Patch, H.M., Biddinger, D.J., Coccia, M., Crone, M.K.,
374 Fiely, M., Francis, J.S., Hines, H.M., and Hodges, M. (2020). Pollen protein: lipid
375 macronutrient ratios may guide broad patterns of bee species floral preferences. *Insects*
376 11, 132.
- 377 9. Paoli, P.P., Donley, D., Stabler, D., Saseendranath, A., Nicolson, S.W., Simpson, S.J.,
378 and Wright, G.A. (2014). Nutritional balance of essential amino acids and
379 carbohydrates of the adult worker honeybee depends on age. *Amino Acids* 46, 1449–
380 1458.
- 381 10. Ruedenauer, F.A., Raubenheimer, D., Kessner-Beierlein, D., Grund-Mueller, N.,
382 Noack, L., Spaethe, J., and Leonhardt, S.D. (2020). Best be(e) on low fat: linking
383 nutrient perception, regulation and fitness. *Ecol. Lett.* 23, 545–554.
- 384 11. Kraus, S., Gómez-Moracho, T., Pasquaretta, C., Latil, G., Dussutour, A., and Lihoreau,
385 M. (2019). Bumblebees adjust protein and lipid collection rules to the presence of
386 brood. *Curr. Zool.* 65, 437–446.
- 387 12. Palmer-Young, E.C., Farrell, I.W., Adler, L.S., Milano, N.J., Egan, P.A., Junker, R.R.,
388 Irwin, R.E., and Stevenson, P.C. (2019). Chemistry of floral rewards: intra- and
389 interspecific variability of nectar and pollen secondary metabolites across taxa. *Ecol.*
390 *Monogr.* 89, e01335.
- 391 13. Barlow, S.E., Wright, G.A., Ma, C., Barberis, M., Farrell, I.W., Marr, E.C., Brankin,
392 A., Pavlik, B.M., and Stevenson, P.C. (2017). Distasteful nectar deters floral robbery.
393 *Curr. Biol.* 27, 2552–2558.
- 394 14. Muth, F., Francis, J.S., and Leonard, A.S. (2016). Bees use the taste of pollen to
395 determine which flowers to visit. *Biol. Lett.* 12, 20160356.
- 396 15. Francis, J.S., Acevedo, C.R., Muth, F., and Leonard, A.S. (2019). Nectar quality
397 changes the ecological costs of chemically defended pollen. *Curr. Biol.* 29, R679–
398 R680.
- 399 16. Sedivy, C., Müller, A., and Dorn, S. (2011). Closely related pollen generalist bees
400 differ in their ability to develop on the same pollen diet: evidence for physiological
401 adaptations to digest pollen. *Funct. Ecol.* 25, 718–725.
- 402 17. Wang, X.-Y., Tang, J., Wu, T., Wu, D., and Huang, S.-Q. (2019). Bumblebee rejection
403 of toxic pollen facilitates pollen transfer. *Curr. Biol.* 29, 1401–1406.

- 404 18. Stevenson, P.C. (2019). For antagonists and mutualists: the paradox of insect toxic
405 secondary metabolites in nectar and pollen. *Phytochem. Rev.*, 1–12.
- 406 19. Rivest, S., and Forrest, J.R.K. (2020). Defence compounds in pollen: why do they
407 occur and how do they affect the ecology and evolution of bees? *New Phytol.* 225,
408 1053–1064.
- 409 20. Wright, G.A., Nicolson, S.W., and Shafir, S. (2018). Nutritional physiology and
410 ecology of honey bees. *Annu. Rev. Entomol.* 63.
- 411 21. Trinkl, M., Kaluza, B.F., Wallace, H., Heard, T.A., Keller, A., and Leonhardt, S.D.
412 (2020). Floral Species Richness Correlates with Changes in the Nutritional Quality of
413 Larval Diets in a Stingless Bee. *Insects* 11, 125.
- 414 22. Lucchetti, M.A., Kilchenmann, V., Glauser, G., Praz, C., and Kast, C. (2018). Nursing
415 protects honeybee larvae from secondary metabolites of pollen. *Proc. R. Soc. B Biol.*
416 *Sci.* 285, 20172849.
- 417 23. Pereboom, J.J.M. (2000). The composition of larval food and the significance of
418 exocrine secretions in the bumblebee *Bombus terrestris*. *Insectes Soc.* 47, 11–20.
- 419 24. Norden, B., Batra, S.W.T., Fales, H.M., Hefetz, A., and Shaw, G.J. (1980).
420 Anthophora bees: unusual glycerides from maternal Dufour's glands serve as larval
421 food and cell lining. *Science* (80-.). 207, 1095–1097.
- 422 25. Voulgari-Kokota, A., McFrederick, Q.S., Steffan-Dewenter, I., and Keller, A. (2019).
423 Drivers, Diversity, and Functions of the Solitary-Bee Microbiota. *Trends Microbiol.*
- 424 26. Eckhardt, M., Haider, M., Dorn, S., and Müller, A. (2014). Pollen mixing in pollen
425 generalist solitary bees: a possible strategy to complement or mitigate unfavourable
426 pollen properties? *J. Anim. Ecol.* 83, 588–597.
- 427 27. Nürnberger, F., Keller, A., Härtel, S., and Steffan-Dewenter, I. (2019). Honey bee
428 waggle dance communication increases diversity of pollen diets in intensively
429 managed agricultural landscapes. *Mol. Ecol.* 28, 3602–3611.
- 430 28. Kaluza, B.F., Wallace, H.M., Heard, T.A., Minden, V., Klein, A., and Leonhardt, S.D.
431 (2018). Social bees are fitter in more biodiverse environments. *Sci. Rep.* 8, 12353.
- 432 29. Kaluza, B.F., Wallace, H., Keller, A., Heard, T.A., Jeffers, B., Drescher, N., Blüthgen,
433 N., and Leonhardt, S.D. (2017). Generalist social bees maximize diversity intake in
434 plant species-rich and resource-abundant environments. *Ecosphere* 8, e01758.
- 435 30. Kim, W., Gilet, T., and Bush, J.W.M. (2011). Optimal concentrations in nectar
436 feeding. *Proc. Natl. Acad. Sci.* 108, 16618–16621.
- 437 31. Wei, J., Huo, Z., Gorb, S.N., Rico-Guevara, A., Wu, Z., and Wu, J. (2020). Sucking or

- 438 lapping: facultative feeding mechanisms in honeybees (*Apis mellifera*). *Biol. Lett.* *16*,
439 20200449.
- 440 32. Pattrick, J.G., Symington, H.A., Federle, W., and Glover, B.J. (2020). The mechanics
441 of nectar offloading in the bumblebee *Bombus terrestris* and implications for optimal
442 concentrations during nectar foraging. *J. R. Soc. Interface* *17*, 20190632.
- 443 33. Minnaar, C., Anderson, B., de Jager, M.L., and Karron, J.D. (2019). Plant–pollinator
444 interactions along the pathway to paternity. *Ann. Bot.* *123*, 225–245.
- 445 34. Castellanos, M.C., Wilson, P., Keller, S.J., Wolfe, A.D., and Thomson, J.D. (2006).
446 Anther evolution: pollen presentation strategies when pollinators differ. *Am. Nat.* *167*,
447 288–296.
- 448 35. Westerkamp, C., and Weber, A. (1999). Keel flowers of the Polygalaceae and
449 Fabaceae: a functional comparison. *Bot. J. Linn. Soc.* *129*, 207–221.
- 450 36. Reith, M., Claßen-Bockhoff, R., and Speck, T. (2006). Biomechanics of *Salvia*
451 flowers: the role of lever and flower tube in specialization on pollinators. In *Ecology*
452 *and biomechanics: a mechanical approach to the ecology of animals and plants* (Boca
453 Raton: CRC Press), pp. 123–146.
- 454 37. De Kock, C., Minnaar, C., Lunau, K., Wester, P., Verhoeven, C., Schulze, M.J.,
455 Randle, M.R., Robson, C., Bolus, R.H., and Anderson, B. (2018). The functional role
456 of the keel crest in *Polygala myrtifolia* (Polygalaceae) and its effects on pollinator
457 visitation success. *South African J. Bot.* *118*, 105–111.
- 458 38. Armbruster, W.S., Edwards, M.E., and Debevec, E.M. (1994). Floral character
459 displacement generates assemblage structure of Western-Australian triggerplants
460 (*Stylidium*). *Ecology* *75*, 315–329.
- 461 39. Siriani-Oliveira, S., Oliveira, R., and Schlindwein, C. (2018). Pollination of
462 *Blumenbachia amana* (Loasaceae): flower morphology and partitioned pollen
463 presentation guarantee a private reward to a specialist pollinator. *Biol. J. Linn. Soc.*
464 *124*, 479–491.
- 465 40. Switzer, C.M., Combes, S.A., and Hopkins, R. (2018). Dispensing Pollen via Catapult:
466 Explosive Pollen Release in Mountain Laurel (*Kalmia latifolia*). *Am. Nat.* *191*, 767–
467 776.
- 468 41. Zhang, Q., Fu, W.-L., Wang, X.-F., and Huang, L.-J. (2020). Ingenious floral structure
469 drives explosive pollination in *Hydrilla verticillata* (Hydrocharitaceae). *Plant Biol.* *22*,
470 480–486.
- 471 42. Vallejo-Marín, M., Manson, J.S., Thomson, J.D., and Barrett, S.C.H. (2009). Division

- 472 of labour within flowers: heteranthery, a floral strategy to reconcile contrasting pollen
473 fates. *J. Evol. Biol.* 22, 828–839.
- 474 43. Vallejo-Marín, M. (2019). Buzz pollination: studying bee vibrations on flowers. *New*
475 *Phytol.* 224, 1068–1074.
- 476 44. Córdoba, S.A., and Cocucci, A.A. (2011). Flower power: its association with bee
477 power and floral functional morphology in papilionate legumes. *Ann. Bot.* 108, 919–
478 931.
- 479 45. Harder, L.D. (1990). Pollen removal by bumble bees and its implications for pollen
480 dispersal. *Ecology* 71, 1110–1125.
- 481 46. Yeo, P.F. (2012). *Secondary pollen presentation: form, function and evolution*
482 (Springer Science & Business Media).
- 483 47. Claßen-Bockhoff, R., Speck, T., Tweraser, E., Wester, P., Thimm, S., and Reith, M.
484 (2004). The staminal lever mechanism in *Salvia L.*(Lamiaceae): a key innovation for
485 adaptive radiation? *Org. Divers. Evol.* 4, 189–205.
- 486 48. Buchmann, S.L. (1983). Buzz pollination in angiosperms. In *Handbook of*
487 *Experimental Pollination Biology*, C. Jones and R. Little, eds. (New York: Van
488 Nostrand Reinhold Company), pp. 73–113.
- 489 49. Harder, L.D., and Barclay, R.M.R. (1994). The functional significance of poricidal
490 anthers and buzz pollination: controlled pollen removal from *Dodecatheon*. *Funct.*
491 *Ecol.* 8, 509–517.
- 492 50. Konzmann, S., Hilgendorf, F., Niester, C., Rech, A.R., and Lunau, K. (2020).
493 Morphological specialization of heterantherous *Rhynchanthera grandiflora*
494 (Melastomataceae) accommodates pollinator diversity. *Plant Biol.*
- 495 51. Cardinal, S., and Danforth, B.N. (2013). Bees diversified in the age of eudicots. *Proc.*
496 *R. Soc. B Biol. Sci.* 280, 20122686–20122686.
- 497 52. Cardinal, S., Buchmann, S.L., and Russell, A.L. (2018). The evolution of floral
498 sonication, a pollen foraging behavior used by bees (*Anthophila*). *Evolution* (N. Y.).
499 72, 590–600.
- 500 53. Pritchard, D.J., and Vallejo-Marín, M. (2020). Buzz pollination. *Curr. Biol.* 30, R858–
501 R860.
- 502 54. De Luca, P.A., and Vallejo-Marín, M. (2013). What’s the ‘buzz’ about? The ecology
503 and evolutionary significance of buzz-pollination. *Curr. Opin. Plant Biol.* 16, 429–435.
- 504 55. Pritchard, D.J., and Vallejo-Marín, M. (2020). Floral vibrations by buzz-pollinating
505 bees achieve higher frequency, velocity and acceleration than flight and defence

- 506 vibrations. *J. Exp. Biol.* 223, jeb.220541.
- 507 56. Greenleaf, S.S., Williams, N.M., Winfree, R., and Kremen, C. (2007). Bee foraging
508 ranges and their relationship to body size. *Oecologia* 153, 589–596.
- 509 57. Irwin, R.E., Bronstein, J.L., Manson, J.S., and Richardson, L. (2010). Nectar robbing:
510 ecological and evolutionary perspectives. *Annu. Rev. Ecol. Evol. Syst.* 41, 271–292.
- 511 58. van der Kooi, C.J., Dyer, A.G., Kevan, P.G., and Lunau, K. (2019). Functional
512 significance of the optical properties of flowers for visual signalling. *Ann. Bot.* 123,
513 263–276.
- 514 59. Raguso, R.A. (2008). Wake up and smell the roses: the ecology and evolution of floral
515 scent. *Annu. Rev. Ecol. Evol. Syst.* 39, 549–569.
- 516 60. Lunau, K., Papiorek, S., Eltz, T., and Sazima, M. (2011). Avoidance of achromatic
517 colours by bees provides a private niche for hummingbirds. *J. Exp. Biol.* 214, 1607–
518 1612.
- 519 61. Briscoe, A.D., and Chittka, L. (2001). The evolution of color vision in insects. *Annu.*
520 *Rev. Entomol.* 46, 471–510.
- 521 62. van der Kooi, C.J., Stavenga, D.G., Belusic, G., Arikawa, K., and Kelber, A. (2021).
522 Evolution of insect colour vision - from spectral sensitivity to visual ecology. *Annu.*
523 *Rev. Entomol.* 66, DOI 10.1146/annurev-ento-0.
- 524 63. Burger, H., Jürgens, A., Ayasse, M., and Johnson, S.D. (2017). Floral signals and
525 filters in a wasp-and a bee-pollinated *Gomphocarpus* species (Apocynaceae:
526 Asclepiadeae). *Flora* 232, 83–91.
- 527 64. Hetherington-Rauth, M.C., and Ramírez, S.R. (2016). Evolution and diversity of floral
528 scent chemistry in the euglossine bee-pollinated orchid genus *Gongora*. *Ann. Bot.* 118,
529 135–148.
- 530 65. Junker, R.R., and Blüthgen, N. (2010). Floral scents repel facultative flower visitors,
531 but attract obligate ones. *Ann. Bot.* 105, 777–782.
- 532 66. Ohashi, K., Makino, T.T., and Arikawa, K. (2015). Floral colour change in the eyes of
533 pollinators: testing possible constraints and correlated evolution. *Funct. Ecol.* 29,
534 1144–1155.
- 535 67. Kemp, J.E., and Ellis, A.G. (2019). Cryptic petal coloration decreases floral apparency
536 and herbivory in nocturnally closing daisies. *Funct. Ecol.* 33, 2130–2141.
- 537 68. Kraaij, M., and van der Kooi, C.J. (2020). Surprising absence of association between
538 flower surface microstructure and pollination system. *Plant Biol.* 22, 177–183.
- 539 69. Stavenga, D.G., Staal, M., and van der Kooi, C.J. (2020). Conical epidermal cells

- 540 cause velvety colouration and enhanced patterning in *Mandevilla* flowers. *Faraday*
541 *Discuss.*, In press.
- 542 70. Chapurlat, E., Anderson, J., Ågren, J., Friberg, M., and Sletvold, N. (2018). Diel
543 pattern of floral scent emission matches the relative importance of diurnal and
544 nocturnal pollinators in populations of *Gymnadenia conopsea*. *Ann. Bot.* *121*, 711–
545 721.
- 546 71. Balducci, M.G., Van der Niet, T., and Johnson, S.D. (2020). Diel scent and nectar
547 rhythms of an African orchid in relation to bimodal activity patterns of hawkmoth
548 pollinators. *Ann. Bot.*
- 549 72. Martignier, T., Labouche, A.-M., and Pannell, J.R. (2019). Pollination elicits an
550 accelerated reduction in nocturnal scent emission by flowers of the dioecious herb
551 *Silene latifolia*. *Botany* *97*, 495–502.
- 552 73. Renner, S.S. (2006). Rewardless flowers in the Angiosperms and the role of insect
553 cognition in their evolution. In *Plant-pollinator interactions: from specialization to*
554 *generalization*, N. M. Waser and J. Ollerton, eds. (Chicago: Princeton University
555 Press).
- 556 74. Minckley, R., and Roulston, T.H. (2006). Incidental mutualisms and pollen
557 specialization among bees. In *Plant-Pollinator Interactions. From specialization to*
558 *generalization*, N. Waser and J. Ollerton, eds. (The University of Chicago Press), p.
559 488.
- 560 75. Schiestl, F.P. (2017). Innate receiver bias: its role in the ecology and evolution of
561 plant–animal interactions. *Annu. Rev. Ecol. Evol. Syst.* *48*, 585–603.
- 562 76. Lunau, K., and Maier, E.J. (1995). Innate colour preferences of flower visitors. *J.*
563 *Comp. Physiol. A* *177*, 1–19.
- 564 77. Kinoshita, M., Stewart, F.J., and Ômura, H. (2017). Multisensory integration in
565 *Lepidoptera*: Insights into flower-visitor interactions. *BioEssays* *39*, 1600086.
- 566 78. Dyer, A.G., Boyd-Gerny, S., Shrestha, M., Garcia, J.E., van der Kooi, C.J., and Wong,
567 B.B.M. (2019). Colour preferences of *Tetragonula carbonaria* stingless bees for colour
568 morphs of the Australian native orchid *Caladenia carnea*. *J. Comp. Physiol. A* *205*,
569 347–361.
- 570 79. Ayasse, M., Schiestl, F.P., Paulus, H.F., Ibarra, F., and Francke, W. (2003). Pollinator
571 attraction in a sexually deceptive orchid by means of unconventional chemicals. *Proc.*
572 *R. Soc. B Biol. Sci.* *270*, 517–522.
- 573 80. van der Kooi, C.J., Kevan, P.G., and Koski, M.H. (2019). The thermal ecology of

- 574 flowers. *Ann. Bot.* *124*, 343–353.
- 575 81. Smithson, A., and Macnair, M.R. (1997). Negative frequency-dependent selection by
576 pollinators on artificial flowers without rewards. *Evolution* (N. Y). *51*, 715–723.
- 577 82. Gigord, L.D.B., Macnair, M.R., and Smithson, A. (2001). Negative frequency-
578 dependent selection maintains a dramatic flower color polymorphism in the rewardless
579 orchid *Dactylorhiza sambucina* (L.) Soo. *Proc. Natl. Acad. Sci.* *98*, 6253–6255.
- 580 83. Stejskal, K., Streinzer, M., Dyer, A., Paulus, H.F., and Spaethe, J. (2015). Functional
581 significance of labellum pattern variation in a sexually deceptive orchid (*Ophrys*
582 *heldreichii*): evidence of individual signature learning effects. *PLoS One* *10*.
- 583 84. Ayasse, M., Schiestl, F.P., Paulus, H.F., Löfstedt, C., Hansson, B., Ibarra, F., and
584 Francke, W. (2000). Evolution of reproductive strategies in the sexually deceptive
585 orchid *Ophrys sphegodes*: how does flower-specific variation of odor signals influence
586 reproductive success? *Evolution* (N. Y). *54*, 1995–2006.
- 587 85. Russell, A.L., Kikuchi, D.W., Giebink, N.W., and Papaj, D.R. (2020). Sensory bias
588 and signal detection trade-offs maintain intersexual floral mimicry. *Philos. Trans. R.*
589 *Soc. B* *375*, 20190469.
- 590 86. Wong, B.B.M., and Schiestl, F.P. (2002). How an orchid harms its pollinator. *Proc. R.*
591 *Soc. B Biol. Sci.* *269*, 1529–1532.
- 592 87. Knauer, A.C., and Schiestl, F.P. (2015). Bees use honest floral signals as indicators of
593 reward when visiting flowers. *Ecol. Lett.* *18*, 135–143.
- 594 88. Burdon, R.C.F., Raguso, R.A., Gegear, R.J., Pierce, E.C., Kessler, A., and
595 Parachnowitsch, A.L. (2020). Scented nectar and the challenge of measuring honest
596 signals in pollination. *J. Ecol.*, In press.
- 597 89. Wright, G.A., and Schiestl, F.P. (2009). The evolution of floral scent: the influence of
598 olfactory learning by insect pollinators on the honest signalling of floral rewards.
599 *Funct. Ecol.* *23*, 841–851.
- 600 90. Leonard, A.S., Dornhaus, A., and Papaj, D.R. (2011). Why are floral signals complex?
601 An outline of functional hypotheses (Cambridge University Press, Cambridge).
- 602 91. Anderson, B., and Johnson, S.D. (2008). The geographical mosaic of coevolution in a
603 plant-pollinator mutualism. *Evolution* (N. Y). *62*, 220–225.
- 604 92. Thompson, J.N., and Cunningham, B.M. (2002). Geographic structure and dynamics
605 of coevolutionary selection. *Nature* *417*, 735–738.
- 606 93. Venjakob, C., Leonhardt, S.D., and Klein, A.-M. (2020). Inter-Individual nectar
607 chemistry changes of field scabious, *Knautia arvensis*. *Insects* *11*, 75.

- 608 94. Bailes, E.J., Pattrick, J.G., and Glover, B.J. (2018). An analysis of the energetic reward
609 offered by field bean (*Vicia faba*) flowers: Nectar, pollen, and operative force. *Ecol.*
610 *Evol.* 8, 3161–3171.
- 611 95. Parachnowitsch, A.L., Manson, J.S., and Sletvold, N. (2019). Evolutionary ecology of
612 nectar. *Ann. Bot.* 123, 247–261.
- 613 96. Nakamura, S., and Kudo, G. (2016). Foraging responses of bumble bees to rewardless
614 floral patches: importance of within-plant variance in nectar presentation. *AoB Plants*
615 8.
- 616 97. Brown, M., Downs, C.T., and Johnson, S.D. (2011). Covariation of flower traits and
617 bird pollinator assemblages among populations of *Kniphofia linearifolia*
618 (*Asphodelaceae*). *Plant Syst. Evol.* 294, 199–206.
- 619 98. Streinzer, M., Roth, N., Paulus, H.F., and Spaethe, J. (2019). Color preference and
620 spatial distribution of glaphyrid beetles suggest a key role in the maintenance of the
621 color polymorphism in the peacock anemone (*Anemone pavonina*, *Ranunculaceae*) in
622 Northern Greece. *J. Comp. Physiol. A* 205, 735–743.
- 623 99. Martínez-Harms, J., Hadar, R., Márquez, N., Menzel, R., Shmida, A., Stavenga, D.G.,
624 and Vorobyev, M. (2020). Enhanced UV-Reflection Facilitated a Shift in the
625 Pollination System of the Red Poppy, *Papaver rhoeas* (*Papaveraceae*). *Plants* 9, 927.
- 626 100. Gross, K., Sun, M., and Schiestl, F.P. (2016). Why do floral perfumes become
627 different? Region-specific selection on floral scent in a terrestrial orchid. *PLoS One*
628 11, e0147975.
- 629 101. Parker, A.J., Williams, N.M., and Thomson, J.D. (2018). Geographic patterns and
630 pollination ecotypes in *Claytonia virginica*. *Evolution (N. Y.)*. 72, 202–210.
- 631 102. van der Kooi, C.J., and Ollerton, J. (2020). The origins of flowering plants and
632 pollinators. *Science (80-.)*. 368, 1306–1308.
- 633 103. Ramírez, S.R., Eltz, T., Fujiwara, M.K., Gerlach, G., Goldman-Huertas, B., Tsutsui,
634 N.D., and Pierce, N.E. (2011). Asynchronous diversification in a specialized plant-
635 pollinator mutualism. *Science (80-.)*. 333, 1742–1746.
- 636 104. van der Niet, T., and Johnson, S.D. (2012). Phylogenetic evidence for pollinator-
637 driven diversification of angiosperms. *Trends Ecol. Evol.* 27, 353–361.
- 638 105. Wang, T.N., Clifford, M.R., Martínez-Gómez, J., Johnson, J.C., Riffell, J.A., and Di
639 Stilio, V.S. (2019). Scent matters: differential contribution of scent to insect response
640 in flowers with insect vs. wind pollination traits. *Ann. Bot.* 123, 289–301.

641

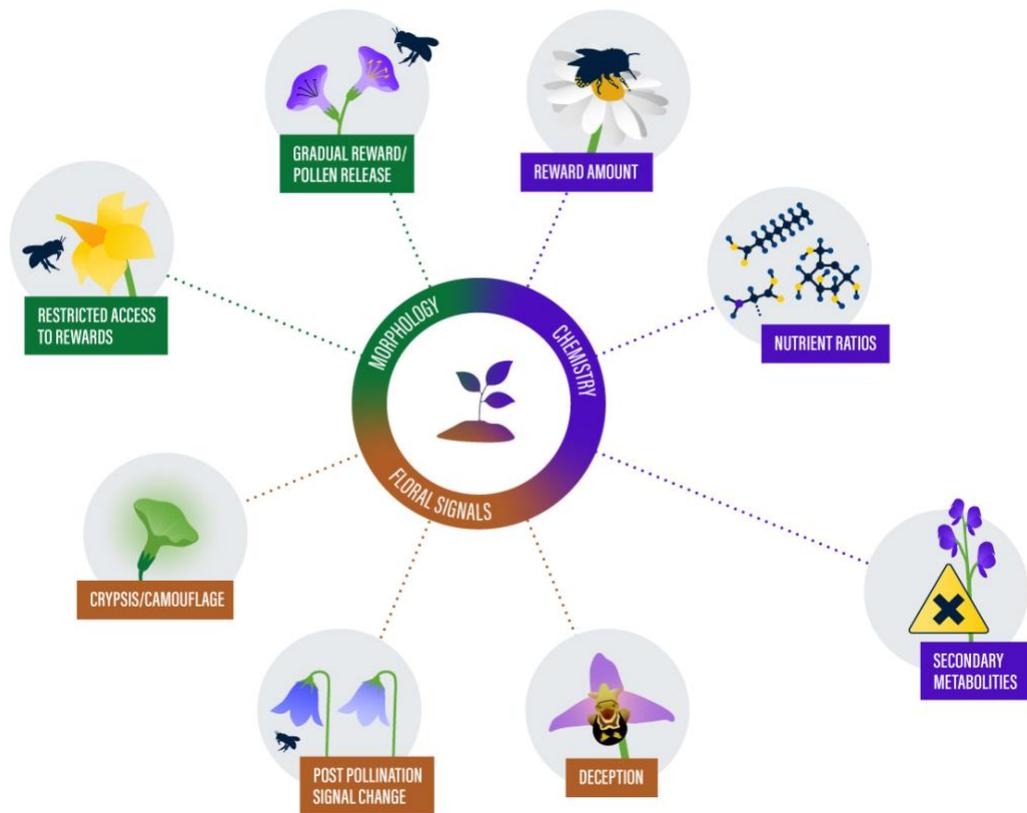
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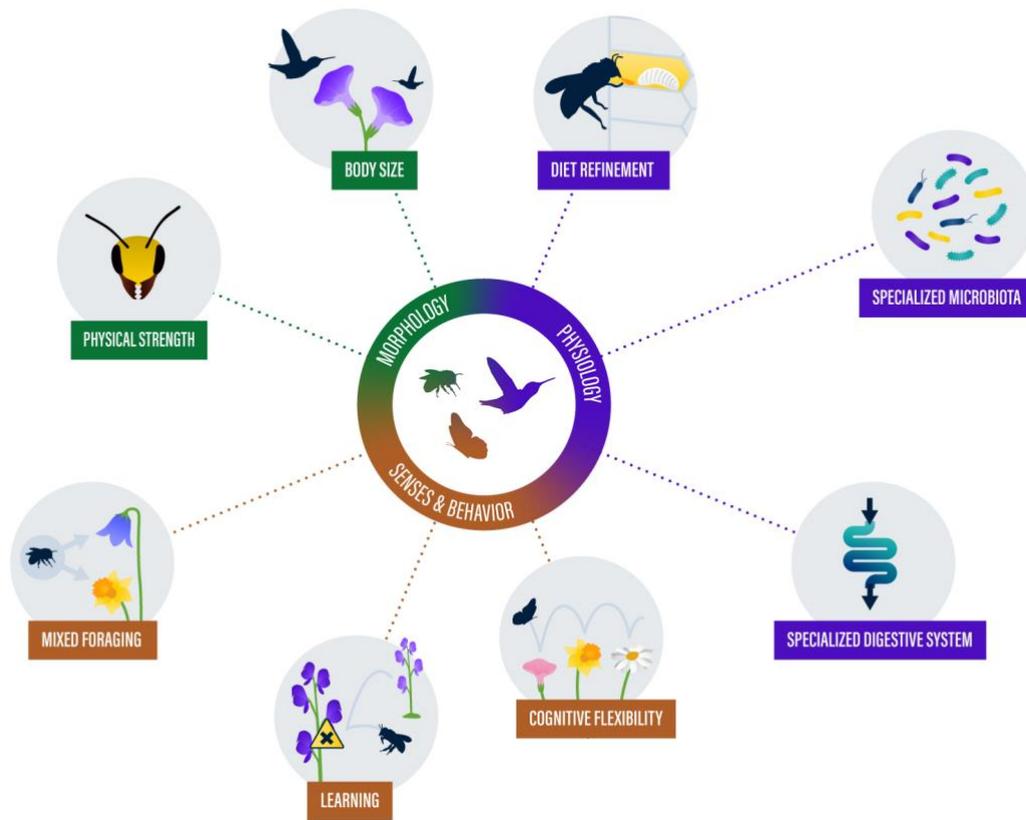
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645 Figure 1. Varying symmetry and relative benefits for flower visitors and plants. The
 646 horizontal bar depicts the relative benefit for pollinator (left) and plant (right). The figure
 647 shows exemplary cases. The position on the continuum strongly depends on the context, and
 648 is determined by the pollinator's relative efficacy (pollen transferred versus reward removed),
 649 and the amount of reward obtained over energy invested.



650

651 Figure 2. Exemplary plant adaptations to increase advantage from interaction with
 652 pollinators. Characteristics of the reward chemistry can render the reward quantity or quality
 653 suboptimal, and thereby deter certain flower visitors or incentivise a return visit. Signals
 654 acting on pollinator senses can increase or decrease floral attraction to pollinators and other
 655 flower visitors. Flower morphology and biomechanics determine the handling time and
 656 accessibility of rewards.



657

658 Figure 3. Exemplary pollinator adaptations and behavioural responses to increase
 659 exploitations of flowers. Pollinator senses and foraging behaviour determine how easily a
 660 pollinator can (learn to) detect or avoid flowers and associate signals with a reward.
 661 Morphological adaptations, such as body size, determine whether a pollinator can reach and
 662 enter the flower and access rewards. Physiological adaptations allow pollinators to efficiently
 663 exploit the rewards and to cope with reward toxicity.