

**ScienceDirect** 

# Engineering plant-based feedstocks for sustainable aquaculture



Johnathan A. Napier<sup>1</sup> and Monica B. Betancor<sup>2</sup>

#### Abstract

There is a growing recognition of the challenges associated with ensuring good nutrition for all without compromising the environment. This is particularly true for aquaculture, given the reliance on marine extraction for key feed ingredients, yet at the same time it delivers key nutrients such as omega-3 long chain polyunsaturated fatty acids. This review will consider progress in transitioning away from oceanic-derived fish oils as feed ingredients, focusing on the emerging transgenic plant sources of these fatty acids. Specific consideration is given to the "validation" phase of this process, in which oils from GM plants are used as substitutes for bona fide fish oils in aguafeed diets. Equally, consideration is given to the demonstration of "real-world" potential by GM field trials. Collectively, the status of these new plant-based sources of omega-3 fish oils confirm the arrival of a new wave of plant biotech products, 25 years after the introduction of herbicide-tolerant input traits and demonstrate the power of GM agriculture to contribute to food security and operating within planetary boundaries.

#### Addresses

<sup>1</sup> Rothamsted Research, Harpenden, Herts AL5 2JQ, UK

<sup>2</sup> Institute of Aquaculture, University of Stirling, Stirling, FK9 4LA, UK

Corresponding author: Napier, Johnathan A. (Johnathan.napier@rothamsted.ac.uk)

#### Current Opinion in Plant Biology 2023, 71:102323

This review comes from a themed issue on Plant Biotechnology (2023)

Edited by Nicola J. Patron and Steven Burgess

For complete overview of the section, please refer the article collection -Plant Biotechnology (2023)

Available online 9 December 2022

https://doi.org/10.1016/j.pbi.2022.102323

1369-5266/© 2022 The Author(s). Published by Elsevier Ltd. This is an open access article under the CC BY license (http://creativecommons. org/licenses/by/4.0/).

#### Keywords

Aquaculture, Plant biotechnology, Omega-3, Fish oils, Sustainability.

### Author declaration

JAN is listed as an inventor on several patents relating to the production of EPA and DHA in transgenic camelina and acts as a Scientific Advisor for Yield10 Biosciences. MBB declares no interest.

### Introduction

The health-benefits associated with a diet containing oily fish rich in omega-3 long chain polyunsaturated fatty acids (omega-3 LC-PUFAs) are now well-established, linking regular consumption of these fatty acids with reduced risk of cardiovascular disease and myocardial infarction [1]. However, many factors have conspired to make this apparently straightforward recommendation less easy to fulfil, including the relentless growth in the global population which puts increasing demands on our planet's finite resources, pollution of the marine environments and advancing climate change [2,3]. Perhaps counterintuitively, the growth in aquaculture as the dominant farming system to produce fish for human consumption, has altered the dynamics of omega-3 supply and demand, putting additional strain on finite global fish stocks [2,3]. As discussed below, efforts to make aquaculture more sustainable have lowered the nutritional value of the final product [4]. In this sense, the inclusion of terrestrial plant oils, the main alternative to fish oil in Europe, in aquafeeds has led to a marked reduction in the levels of omega-3 LC-PUFAs accompanied by increased levels of short chain fatty acids characteristic of terrestrial systems. Long established dietary patterns have radically shifted in the last few decades, to the point that metabolic diseases resulting from the overconsumption of calorie-rich but low nutrient diets outweighs (no pun intended) the number of people suffering from malnutrition due to insufficient food. Simultaneous to that, a growing awareness that global patterns of food consumption are exceeding planetary boundaries has also led for calls in a radical shift in how we feed ourselves, most notably with a strong decrease in the consumption of (terrestrial) animal protein and concomitant increase in dietary plant protein [5]. Interestingly, elevated consumption of fish is also recommended (because of the nutritional benefits of omega-3s etc) but there is an inherent tension associated with such advice, not least of all with respect to sustainability of global fish stocks [2,5,6,7].

Plant biotechnology has always aspired to deliver benefits to society, even if this has sometimes not been obvious [8]. The first wave of GM crops developed in the 1990s was focussed on delivering so-called input traits, which help farmers (via herbicide tolerance and/or herbivore resistance) but provide no direct benefit to the consumer or nutritional enhancement (although they do deliver a reduced reliance on chemical inputs). A second wave of output traits for improved nutrient content is only now starting to appear, 25 years behind the input traits – the many and varied reasons for this have been recently considered by us and other [8,9]. At the forefront of these new GM crops are plants which have been engineered to accumulated omega-3 LC-PUFAs in their seed oils, providing a novel terrestrial source of this important fatty acids independent of marine stocks [10] – such plant-based sources of "omega-3 fish oils" can be used for both direct human nutrition or as animal feed ingredients (in either aquaculture or terrestrial animals), delivering benefit by these different routes [11].

The omega-3 LC-PUFAs, eicosapentaenoic acid (EPA;  $20:5\Delta^{5,8,11,14,17}$ ) and docosahexaenoic acid (DHA;  $22:6\Delta^{4,7,10,13,16,19}$ ) are found in fish oils and are almost exclusively restricted to aquatic foodwebs [2,3]. The predominant biosynthesisers of EPA and DHA in the marine ecosystems are microalgae and other phytoplankton, with the fatty acids accumulating in the multiple trophic levels above these founders [2]. Marine and salmonid fish species have little or no endogenous capacity to synthesise these fatty acids, relying instead on dietary intake. No angiosperm plant species have the capacity to synthesise EPA and DHA, although C18 precursor polyunsaturated fatty acids are ubiquitous in the Plant Kingdom [4]. Since the 1990s, significant efforts have been made towards engineering plants to accumulate EPA and/or DHA as an alternative to marine extraction [8-10]. Representing one of the very few projects to progress first from a discovery phase into application, the production of omega-3 fish oils in transgenic plants represents a significant achievement for plant biotechnology. This short review will concentrate on recent examples of scale-up and translation, including demonstration of efficacy in aquafeed trials. Aspect of the molecular biology and genetic toolkits that have been used to generate these transgenic plants have been recently considered elsewhere and are therefore only discussed briefly.

# Transgenic plants as a platform for the synthesis of EPA + DHA

Despite being a deceptively straightforward objective, the efficient synthesis of the non-native fatty acids EPA and DHA proved challenging to achieve, especially when accumulation was restricted to storage triacylglycerols via seed-specific promoters [12-14]. This was due in part to the relative complexity of the biosynthetic pathway, requiring the co-ordinated expression of a minimum of five heterologous genes, representing some of the most complex metabolic engineering attempted in transgenic plants [10]. However, by adopting an iterative approach to defining the optimal combination of genes and regulatory elements, significant progress has been achieved in two host species. Camelina [12,15,16] and Canola [17,18]. Interestingly, it has also become apparent that endogenous metabolism plays a key role in enabling the effective reconstitution of the omega-3 LC-PUFA biosynthetic pathway and this inherent capability varies between species. For example, the successful synthesis of EPA + DHA in Arabidopsis or Camelina via the so-called alternative pathway [19,20] was not recapitulated in peanut [21]. Even in the case of effective oilseed chasses such as Canola and Camelina, there are significant differences between the species – for example, a very similar suite of genes and regulatory elements was used to direct the synthesis of EPA + DHA, resulting in  $\sim 20\%$ EPA + DHA in Camelina but only 8% EPA and 1% DHA in Canola. This may indicate additional levels of control, which could include transgene integration site, post-transcriptional regulation, enzyme turnover and substrate channelling. Irrespective of these open questions, it is clear that Brassicacea oilseeds are excellent hosts for the transgenic accumulation of omega-3 LC-PUFAs (summarised in Table 1) and the oils derived from these crops have been used in multiple aquafeed trials. Importantly, the real-world performance of these crops has been demonstrated by multi-location field trials across different continents [22-24]. Recently, gene-editing has been used in conjunction with transgenesis to enhance the accumulation of EPA + DHA in Camelina via the inactivation of a competing endogenous pathway [25].

# Novel oils in aquafeed – validation in fish feeding trials

One key aspect of achieving the successful translation of the biotech studies described above is to validate these novel oils as effective substitutes for fish oils in the diets used in aquaculture (represented in Fig. 1). To date, the majority of research in fish using GM-derived oils rich in n-3 LC-PUFA has predominantly focussed on Atlantic salmon (*Salmo salar*). This is largely due to the fact that salmonids are the primary consumer of extracted fish oils given the energy dense feeds required for farming, as well as its high-end value [4,26]. Therefore, it is of paramount importance that the potential effects of using such substitutes are investigated. Consequently, oils derived from both omega-3 rich Camelina [12,23] and Canola (Event NS-B50027-4) [17,18] have been successfully trialled in several life stages. Freshwater fingerlings fed either 4 or 8% high n-3 GM Canola oil (9.3% omega LC-PUFA) exhibited similar growth and survival to those fed a diet containing fish oil as the main dietary lipid source when cultivated at either 12 or 16 °C [27]. In seawater, no difference in growth were found in smolts fed a Camelina oil rich in either EPA or EPA + DHA (23.9% and 16.7% omega-3 LC-PUFA, respectively) compared to those fed a fish oil rich diet [28–31]. Moreover, all the aforementioned trials showed that fish fed the GM-derived oils had enhanced

#### Table 1

Summary of GM Brassicaceae crops successfully engineered to accumulate EPA and/or DHA. The levels of individual fatty acids are given, along with total amounts of omega-3 LC-PUFAs. The bulk of all successful engineering rationales have used the aerobic desaturase and elongase pathway, although one example is listed using an anaerobic polyketide synthase-like system derived from microbial sources. Please see [2] and [10] for further details of the pathways.

| Crop                           | Pathway   | Omega-3 profile                                       | Total EPA + DHA    | Commercialization status  | Reference            |
|--------------------------------|---|---|--------------------|---|----------------------|
| Camelina<br>Camelina<br>Canola | Aerobic pathway - Rothamsted<br>Aerobic pathway - CSIRO<br>Aerobic pathway - BASF | 10% EPA, 10% DHA<br>2% EPA, 12% DHA<br>7% EPA, 1% DHA | 20%<br>14%<br><10% | Pre-regulatory phase<br>Pre-regulatory phase<br>Approved for commercial | [12]<br>[15]<br>[18] |
| Canola                         | Aerobic pathway - Nuseed/CSIRO  | 1% EPA, 10% DHA                                       | 12%                | growing<br>Approved for commercial<br>growing                           | [17•]                |
| Canola                         | Anaerobic pathway (PKS-like) –<br>Dow Agrosciences                                | 4% DHA  | <5%                | Unknown   | [38]                 |

#### Figure 1



Schematic representation of the requirement for omega-3 fish oils in marine aquaculture and how GM plant-derived fatty acids can be used as a substitute for optimal fish and human nutrition.

levels of n-3 LC-PUFA in their flesh compared to either a commercial diet control or a feed containing wild-type Camelina, while also decreasing the omega-6:omega-3 ratio. For example, salmon fed a diet containing high omega-3 LC-PUFA Camelina oil accumulated almost double the amount of these health beneficial fatty acids as salmon fed a fish oil diet [31]. Evidently, these enhanced levels can contribute to bridging the gap between demand and supply and help meet the 500 mg/ day intake requirement for omega-3 LC-PUFA in order to attain optimal cardiac health [1,2,4]. Enhanced flesh levels of EPA + DHA has been a common finding in other studies where teleost fish were fed omega-3 LC-PUFA vegetable oils such as other fatty species including rainbow trout (Oncorhynchus mykiss) [32], but also gilthead sea bream (Sparus aurata) [33] and sea bass (Dicentrarchus labrax) [34]. In addition to the benefits that the use of these oils can have for consumers, the use of vegetable oils rich in omega-3 LC-PUFA can also have added benefits for the fish such as reduced liver fat infiltration and hepatocyte area relative to fish fed feeds containing wildtype Camelina [35]. Perhaps, most importantly to note is that none of the trials performed to date have reported any adverse effect in fish fed these novel oils thus demonstrating the potential for the exploitation of this key raw material [27-30,31-36]. Studies so far have focussed on the use of the oil, whereas the seed cake has yet to be tested as an alternative protein source. Additional processing of the seeds such as dehulling or solvent extraction might be required to concentrate the protein fraction which in turn will increase the price as well as the environmental footprint of the product.

## Next steps and new approaches

The validation of these novel oils as safe and effective substitutes for oceanic-derived fish oils was a key step in derisking this technology and helping it advance to market. In addition, increased awareness of issues associated with sustainability and environmental footprint [5,11,26] have added impetus to commercialisation. Excitingly, two Canola events developed by BASF/ Cargill and Nuseed/CSIRO have been deregulated by USDA and now can be cultivated at scale [18]. These events represent the most complex metabolic engineering in plants to have been approved for commercial cultivation to date, although these developments represent the culmination of 25 years of fundamental research followed by a concerted focus on development [8]. Simultaneously, oil from the lead Camelina event has been used extensively in human studies, confirming the utility of this oil in both indirect (i.e., animal) and direct human nutrition [37]. There are also efforts to combine a number of different traits to produce a multifunctional crop tailored to the needs of e.g. aquaculture [11], combining the EPA + DHA trait with other high value components of aquafeed diets such as

the antioxidant and pigment astaxanthin, which can be made in a number of different transgenic hosts and has been independently validated as a novel ingredient in fish feeding trials [39,40]. Similarly, the amino acid composition and seed storage proteins can be enhanced to make the resulting protein composition more suitable for fish nutrition (e.g. by the inclusion of the cysteine derivative, taurine) as has been achieved in transgenic soybean [40]. Equally importantly, it is possible to use gene editing to remove compounds such as glucosinolates which can act as feeding deterrents for some fish species [41,42]. On this basis it is possible to envisage a bespoke crop ideotype specifically designed to meet to the nutritional needs of both the fish and the human consumer whilst also delivering to the requirements of operating within planetary boundaries and leaving a lighter environmental footprint on our planet [11,43].

# Conclusions

Recent analysis confirms the significant variance between demand for omega-3 long chain PUFAs and the available supplies from our oceans, currently estimated at providing only  $\sim 30\%$  of the recommended levels of EPA and DHA for the global population [44]. This highlights the pressing need for alternative supplies of these important fatty acids, not least of all given their health-protective role against cardiovascular disease and other metabolic pathologies which now blight the lives of many millions of people. In addition to the sustainable synthesis of EPA & DHA via transgenic plants, a number of microbial (predominantly algae) systems are being established [reviewed in [45]] and also following the same path of validating these novel oils via aquafeed trials [46]. One advantage of using algal-derived omega-3 PUFAs is that these strains are usually not GMOs, making their regulatory approval and consumer acceptance relatively straightforward. However, the largescale culture of algal strains remains a challenge, in terms of economics and infrastructure. Moreover, almost all commercial algal platforms rely on heterotrophic growth (i.e., cultured using an exogenous carbon source which itself is invariably plant-derived, such as sugarcane) - this is not always fully reflected in claims of sustainability, and it would be interesting to carry out a life cycle analysis of the different production systems to allow the consumer to make an informed choice with regard to environmental footprint. Other sources of LC-PUFA such as krill oil are now mainstream ingredients in aquafeed, although their inclusion is normally limited mainly due to its high price [2]. Another approach to boost the levels of these health-beneficial fatty acids in fish without impacting on fisheries is the use of finishing diets high in LC-PUFA [47]. Although the novel plant platforms described above are GM with the associated regulatory burdens [8,9], they have the advantage of utilising the pre-existing infrastructure and know-how which underpin modern agriculture, allowing rapid

scaling and incorporation into production cycles. Expanding our use of plant-derived feed and foodstuffs will be essential for operating within planetary boundaries and collectively we need to embrace a different approach to ensuring optimal nutrition for all - plant biotechnology can play a key role in enabling this. As such, it is important to consider all ethical aspects of using GM-derived ingredients, including the impact of not adopting these innovations [48].

#### Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

#### Data availability

No data was used for the research described in the article.

#### **Acknowledgments**

JAN and MMB gratefully acknowledge support from BBSRC, (UKRI), specifically grants BB/S006400/1 and BB/S005919/1.

#### References

Papers of particular interest, published within the period of review, have been highlighted as:

- of special interest
- •• of outstanding interest
- Innes JK, Calder PC: Marine omega-3 (N-3) fatty acids for 1. cardiovascular health: an update for 2020. Int J Mol Sci 2020. 21:1362. https://doi.org/10.3390/ijms21041362
- Tocher DR, Betancor MB, Sprague M, Olsen RE, Napier JA: 2
- Omega-3 long-chain polyunsaturated fatty acids, EPA and DHA: bridging the gap between supply and demand. Nutrients 2019, 11:89, https://doi.org/10.3390/nu11010089

A comprehensive review comparing different new sources of EPA and DHA, including algal and transgenic plants

- Nichols PD, McManus A, Krail K, Sinclair AJ, Miller M: Recent 3. advances in omega-3: health benefits, sources, products and bioavailability. Nutrients 2014, 6:3727-3733, https://doi.org 10.3390/nu6093727.
- Sprague M, Dick JR, Tocher DR: Impact of sustainable feeds 4. on omega-3 long-chain fatty acid levels in farmed Atlantic salmon, 2006–2015. Sci Rep 2016, 6, 21892.
- Willett W, Rockström J, Loken B, Springmann M, Lang T, 5
- Vermeulen S, ... Murray CJ: Food in the Anthropocene: the EAT–Lancet Commission on healthy diets from sustainable food systems. *Lancet* 2019, **393**:447–492

A ground-breaking analysis of the challenges of feeding the global population within so-called planetary boundaries. The study recommends a shift to a diet which is predominantly plant-based.

- Froehlich HE, Jacobsen NS, Essington TE, et al.: Avoiding the 6. ecological limits of forage fish for fed aquaculture. Nat Sustain 2018, 1:298-303, https://doi.org/10.1038/s41893-018-0077-1
- Cottrell RS, Blanchard JL, Halpern BS, et al.: Global adoption of 7. novel aquaculture feeds could substantially reduce forage fish demand by 2030. Nat Food 2020, 1:301-308, https:// doi.org/10.1038/s43016-020-0078-x.

- Napier JA, Haslam RP, Tsalavouta M, Sayanova O: The chal-8. lenges of delivering genetically modified crops with nutritional enhancement traits. Native Plants 2019, 5:563-567, https://doi.org/10.1038/s41477-019-0430-z
- Van Der Straeten D, Bhullar NK, De Steur H, et al.: Multiplying the efficiency and impact of biofortification through metabolic engineering. Nat Commun 2020, 11:5203, https://doi.org/ 10.1038/s41467-020-19020-4
- 10. Napier JA, Usher S, Haslam RP, Ruiz-Lopez N, Sayanova O: Transgenic plants as a sustainable, terrestrial source of fish oils. Eur J Lipid Sci Technol 2015, 117:1317–1324, https:// doi.org/10.1002/ejlt.201400452.
- 11. Napier JA, Haslam RP, Olsen RE, et al.: Agriculture can help aquaculture become greener. Nat Food 2020, 1:680–683, https://doi.org/10.1038/s43016-020-00182-9.

This review defines the issues facing aquaculture, balancing the laudable goal of becoming more sustainable against the challenge of maintaining the nutritional benefits from consuming fish. A wider role for agriculture and plant biotechnology is proposed.

- Ruiz-Lopez N, Haslam RP, Napier JA, Sayanova O: Successful high-level accumulation of fish oil omega-3 longchain polyunsaturated fatty acids in a transgenic oilseed crop. Plant J 2014, 77:198-208, https://doi.org/10.1111 tpj.12378.
- 13. Petrie JR, Shrestha P, Zhou XR, Mansour MP, Liu Q, Belide S, Nichols PD, Singh SP: Metabolic engineering plant seeds with fish oil-like levels of DHA. PLoS One 2012, 7, e49165, https:// doi.org/10.1371/journal.pone.0049165.
- 14. Haslam RP, Sayanova O, Kim HJ, Cahoon EB, Napier JA: **Synthetic redesign of plant lipid metabolism**. *Plant J* 2016, **87**: 76-86, https://doi.org/10.1111/tpj.13172.
- 15. Petrie JR, Shrestha P, Belide S, Kennedy Y, Lester G, Liu Q, Divi UK, Mulder RJ, Mansour MP, Nichols PD, Singh SP: Metabolic engineering Camelina sativa with fish oil-like levels of DHA. PLoS One 2014, 9, e85061, https://doi.org/10.1371/ journal.pone.0085061
- 16. Mansour MP, Shrestha P, Belide S, Petrie JR, Nichols PD, Singh SP: Characterization of oilseed lipids from "DHA-producing Camelina sativa": a new transformed land plant containing long-chain omega-3 oils. Nutrients 2014, 6: 776-789. https://doi.org/10.3390/nu6020776.
- Petrie JR, Zhou XR, Leonforte A, McAllister J, Shrestha P,
  Kennedy Y, Belide S, Buzza G, Gororo N, Gao W, Lester G, Mansour MP, Mulder RJ, Liu Q, Tian L, Silva C, Cogan N, Nichols PD, Green AG, de Feyter R, ... Singh SP: Development of a Brassica napus (canola) crop containing fish oil-like levels of DHA in the seed oil. Front Plant Sci 2020, 11:727, https://doi.org/10.3389/fpls.2020.00727

This paper describes the molecular and agronomic performance of the specific Canola event taken through the regulatory approval process.

- 18. Napier JA, Olsen RE, Tocher DR: Update on GM canola crops as novel sources of omega-3 fish oils. Plant Biotechnol J 2019, 17:703-705, https://doi.org/10.1111/pbi.13045.
- Qi B, Fraser T, Mugford S, Dobson G, Sayanova O, Butler J, 19. Napier JA, Stobart AK, Lazarus CM: Production of very long chain polyunsaturated omega-3 and omega-6 fatty acids in plants. Nat Biotechnol 2004, 22:739-745, https://doi.org/ 10.1038/nbt972
- 20. Ruiz-Lopez N, Haslam RP, Usher S, Napier JA, Sayanova O: An alternative pathway for the effective production of the omega-3 long-chain polyunsaturates EPA and ETA in transgenic oilseeds. Plant Biotechnology Journal 2015, 13:1264-1275, https://doi.org/10.1111/pbi.12328. 2015.
- 21. Wang C, Qing X, Yu M, Sun Q, Liu F, Qi B, Li X: Production of eicosapentaenoic acid (EPA, 20:5n-3) in transgenic peanut (Arachis hypogaea L.) through the alternative  $\Delta$ 8-desaturase pathway. Mol Biol Rep 2019, 46:333–342, https://doi.org 10.1007/s11033-018-4476-1.

- 22. Usher S, Han L, Haslam RP, Michaelson LV, Sturtevant D, Aziz M, Chapman KD, Sayanova O, Napier JA: Tailoring seed oil composition in the real world: optimising omega-3 long chain polyunsaturated fatty acid accumulation in transgenic Camelina sativa. Sci Rep 2017, 7:6570, https://doi.org/10.1038/ s41598-017-06838-0.
- 23. Han L, Usher S, Sandgrind S, Hassall K, Sayanova O, Michaelson LV, Haslam RP, Napier JA: High level accumulation of EPA and DHA in field-grown transgenic Camelina - a multiterritory evaluation of TAG accumulation and heterogeneity. Plant Biotechnology Journal 2020, 18:2280-2291, https://doi.org/ 10.1111/pbi.1338
- 24. Han L, Silvestre S, Sayanova O, Haslam RP, Napier JA: Using field evaluation and systematic iteration to rationalize the accumulation of omega-3 long-chain polyunsaturated fatty acids in transgenic Camelina sativa. Plant Biotechnology Journal 2022, https://doi.org/10.1111/pbi.13867.

A significant study incorporating field release to determine the perfor-mance of new iterations of transgenic Camelina making EPA and DHA. The need to include real-world performance in the "design-build-testlearn" cycle is emphasised.

- Han L, Haslam RP, Silvestre S, Lu C, Napier JA: Enhancing the 25. accumulation of eicosapentaenoic acid and docosahexae
- noic acid in transgenic Camelina through the CRISPR-Cas9 inactivation of the competing FAE1 pathway. Plant Biotechnology Journal 2022, https://doi.org/10.1111/pbi.13876

The first example of a GM and GE stack undergoing field release. Specifically, gene editing was used to remove a competing pathway, enhancing the transgenic accumulation of EPA and DHA in Camelina.

- Naylor RL, Hardy RW, Buschmann AH, Bush SR, Cao L, 26. Klinger DH, Little DC, Lubchenco J, Shumway SE, Troell M: A 20year retrospective review of global aquaculture. *Nature* 2021, **591**:551–563, https://doi.org/10.1038/s41586-021-03308-6.
- 27.
- Ruyter B, Sissener NH, Østbye TK, Simon CJ, Krasnov A, Bou M, Sanden M, Nichols PD, Lutfi E, Berge GM: n-3 Canola oil effectively replaces fish oil as a new safe dietary source of DHA in feed for juvenile Atlantic salmon. Br J Nutr 2019, 122: 1329–1345, https://doi.org/10.1017/S0007114519002356.

The first study on the performance of the Nuseed DHA Canola oil in aquafeed studies on salmon.

- Betancor MB, Sprague M, Usher S, Sayanova O, Campbell PJ, Napier JA, Tocher DR: A nutritionally-enhanced oil from 28. transgenic Camelina sativa effectively replaces fish oil as a source of eicosapentaenoic acid for fish. Sci Rep 2015, 5: 8104, https://doi.org/10.1038/srep08104.
- 29. Betancor MB, Sprague M, Sayanova O, Usher S, Campbell PJ Napier JA, Caballero MJ, Tocher DR: Evaluation of a high-EPA oil from transgenic Camelina sativa in feeds for Atlantic salmon (Salmo salar L.): effects on tissue fatty acid composition, histology and gene expression. Aquaculture 2015, 444: 1-12, https://doi.org/10.1016/j.aquaculture.2015.03.020
- Betancor MB, Sprague M, Sayanova O, Usher S, Metochis C, Campbell PJ, Napier JA, Tocher DR: Nutritional evaluation of an EPA-DHA oil from transgenic camelina sativa in feeds for post-smolt atlantic salmon (Salmo salar L.). PLoS One 2016, 11, e0159934, https://doi.org/10.1371/journal.pone.015993
- Betancor MB, Li K, Sprague M, Bardal T, Sayanova O, Usher S, Han L, Måsøval K, Torrissen O, Napier JA, Tocher DR, Olsen RE: An oil containing EPA and DHA from transgenic Camelina sativa to replace marine fish oil in feeds for Atlantic salmon (Salmo salar L.): effects on intestinal transcriptome, histology, tissue fatty acid profiles and plasma biochemistry. PLoS One 2017, 12, e0175415, https://doi.org/10.1371/ journal.pone.0175415.
- Osmond AT, Arts MT, Bazinet RP, Napier JA, Han L, Colombo SM: Transgenic camelina oil is an effective source of eicosapentaenoic acid and docosahexaenoic acid in diets for farmed rainbow trout, in terms of growth, tissue fatty acid content, and fillet sensory properties. *J World Aquacult Soc* 2021, **52**:961–986.
- Betancor MB, Sprague M, Montero D, Usher S, Sayanova O, Campbell PJ, Napier JA, Caballero MJ, Izquierdo M, Tocher DR: 33.

Replacement of Marine Fish Oil with de novo Omega-3 Oils from Transgenic Camelina sativa in Feeds for Gilthead Sea Bream (Sparus aurata L.). Lipids 2016, 51:1171-1191, https:// doi.org/10.1007/s11745-016-4191-4

- Betancor MB, MacEwan A, Sprague M, Gong X, Montero D, Han L, Napier JA, Norambuena F, Izquierdo M, Tocher DR: Oil from transgenic Camelina sativa as a source of EPA and DHA in feed for European sea bass (Dicentrarchus labrax L.). Aquaculture 2021, 530, 735759, https://doi.org/10.1016/ j.aquaculture.2020.735759
- Betancor MB, Li K, Bucerzan VS, Sprague M, Sayanova O, Usher S, Han L, Norambuena F, Torrissen O, Napier JA, Tocher DR, Olsen RE: Oil from transgenic Camelina sativa containing over 25 % n-3 long-chain PUFA as the major lipid source in feed for Atlantic salmon (Salmo salar). Br J Nutr 2018, 119:1378-1392, https://doi.org/10.1017/ S0007 114518001125
- 36. Hong J, Bledsoe JW, Overturf KE, Lee S, Iassonova D, Small BC: LatitudeTM oil as a sustainable alternative to dietary fish oil in rainbow trout (Oncorhynchus mykiss): effects on filet fatty acid profiles, intestinal histology, and plasma biochemistry Front Sustain Food Syst 2022, 62

The first study on the performance of the Cargill/BASF EPA Canola oil in aquafeed studies on salmon.

- 37
- West AL, Miles EA, Lillycrop KA, Napier JA, Calder PC, Burdge GC: Genetically modified plants are an alternative to oily fish for providing n-3 polyunsaturated fatty acids in the human diet: a summary of the findings of a Biotech-nology and Biological Sciences Research Council funded project. Nutr Bull 2021, 46:60-68, https://doi.org/10.1111/ nbu 12478

An important demonstration of the utility of EPA + DHA Camelina oil for direct human nutrition. This confirms the ability of this novel oil to substitute for fish oils in direct human nutrition.

- Walsh TA, Bevan SA, Gachotte DJ, et al.: Canola engineered 38 with a microalgal polyketide synthase-like system produces oil enriched in docosahexaenoic acid. Nat Biotechnol 2016, 34: 881-887. https://doi.org/10.1038/nbt.3585
- Nogueira M, Enfissi E, Martínez Valenzuela ME, Menard GN, Driller RL, Eastmond PJ, Schuch W, Sandmann G, Fraser PD: Engineering of tomato for the sustainable production of ketocarotenoids and its evaluation in aquaculture feed. Proc Natl Acad Sci U S A 2017, 114, 10876, https://doi.org/10.1073/ pnas.1708349114. -10881.
- 40. Park H, Weier S, Razvi F, Peña PA, Sims NA, Lowell J, Hungate C, Kissinger K, Key G, Fraser P, Napier JA, Cahoon EB, Clemente TE: Towards the development of a sustainable soya bean-based feedstock for aquaculture. Plant Biotechnology Journal 2017, 15:227-236, https://doi.org/10.1111/pbi.12608
- Neequaye M, Stavnstrup S, Harwood W, Lawrenson T,
  Hundleby P, Irwin J, Troncoso-Rey P, Saha S, Traka MH, Mithen R, Østergaard L: CRISPR-Cas9-Mediated gene editing of MYB28 genes impair glucoraphanin accumulation of Brassica oleracea in the field. *The CRISPR journal* 2021, 4: 416–426, https://doi.org/10.1089/crispr.2021.0007.

Field-based demonstration of trait improvement by gene editing

- 42. Hölzl G, Rezaeva BR, Kumlehn J, Dörmann P: Ablation of glucosinolate accumulation in the oil crop Camelina sativa by targeted mutagenesis of genes encoding the transporters GTR1 and GTR2 and regulators of biosynthesis MYB28 and MYB29. Plant Biotechnol J 2022, https://doi.org/10.1111/ pbi.13936.
- 43. Napier JA, Sayanova O: Nutritional enhancement in plants green and greener. Curr Opin Biotechnol 2020, 61:122-127, https://doi.org/10.1016/j.copbio.2019.12.010
- 44. Hamilton HA, Newton R, Auchterlonie NA, et al.: Systems
- approach to quantify the global omega-3 fatty acid cycle. Nat

Food 2020, 1:59–62, https://doi.org/10.1038/s43016-019-0006-0. An important framework for considering the volume of omega-3 fish oils available to the growing population and how best to close the gap between supply and demand. Current supplies can only provide <40% of demand.

- 45. Sprague M, Betancor MB, Tocher DR: Microbial and genetically engineered oils as replacements for fish oil in aquaculture feeds. *Biotechnol Lett* 2017, **39**:1599–1609, https://doi.org/ 10.1007/s10529-017-2402-6.
- 46. Santigosa E, Brambilla F, Milanese L: Microalgae oil as an effective alternative source of EPA and DHA for gilthead seabream (Sparus aurata) aquaculture. *Animals* 2021, 11:971, https://doi.org/10.3390/ani11040971. Published 2021 Mar 31.
- Xu H, Turchini GM, Francis DS, Liang M, Mock TS, Rombenso A, Ai Q: Are fish what they eat? A fatty acid's perspective. Prog Lipid Res 2020, 80, 101064.
- Hansson SO: A science-informed ethics for agricultural biotechnology. Crop Breeding, Genetics and Genomics 2019 Jul 19, 1.